

NEUROTENSİN: A CENTRAL NERVOUS SYSTEM PEPTIDE

Gülberk UÇAR (*)

Summary: *Neurotensin (NT) is a tridecapeptide mainly distributed in specific areas of the brain and gastrointestinal (GI) tract of many mammalian species. It acts as a local hormone in GI tract mediating the GI motility and secretion of insulin and glucagon; as a neurotransmitter or neuromodulator in the brain effecting cholinergic receptor activation and opioid peptide and catecholamine metabolism. These functions are initiated by specific NT receptors interacted with G proteins and are involved in modulation of second messengers as cAMP, cGMP and inositol phosphates.*

Keywords : *Neurotensin, Neurotensin binding sites, Neurotensin receptor, Central nervous system*

NÖROTENSİN: BİR MERKEZİ SİNİR SİSTEMİ PEPTİDİ

Özet: *Nörotensin (NT), birçok memeli türünün beyin ve mide barsak sisteminin özgül bölgelerinde bulunan 13 amino asitlik bir peptittir. Bu peptit mide-barsak sisteminde yerel hormon olarak görev yapıp mide-barsak hareketlerini ve insülin ve glukagonun salgılanışını düzenlemekte; beyinde sinirsel iletili olarak görev yapıp kolinerjik reseptör aktivasyonu ve opioid peptit, katekolamin metabolizmasını etkilemektedir. Bu işlevler, G proteinleri ile etkileşen ve inositol fosfatlar, cAMP ve cGMP gibi ikincil habercilerin modülasyonunu sağlayan özgül NT reseptörleri tarafından başlatılmaktadır.*

Anahtar Kelimeler : *Nörotensin, Nörotensin bağlanma yeri, Nörotensin reseptörleri, Santral sinir sistemi*

Başvuru Tarihi : 17.1.1992

Kabul Tarihi : 26.3.1992

(*) Hacettepe University, Faculty of Pharmacy, Department of Biochemistry, ANKARA.

Neurotensin (NT) is a 13 amino acid peptide originally isolated and sequenced first from bovine hypothalamus and subsequently from both bovine and human small intestine (1-4). Since its discovery and synthesis, several biological functions have been attributed to this peptide. In this review, the isolation and purification of neurotensin, its structure and distribution in mammalian tissues, NT binding sites and apparent biological actions were summarized.

ISOLATION

NT was discovered in the process of purifying substance P, a hypothalamic peptide, by ion exchange chromatography and clearly separated from the sialogogic activity as a separate peak causing vasodilatation and cyanosis of exposed skin surfaces (1). Using the vasodilatory response to monitor purification procedures, Carraway and Leeman isolated NT and determined its amino acid sequence to be Glu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu-OH (1, 2). Using a purification scheme involving gel filtration, ion exchange chromatography, and high voltage paper electrophoresis, the peptidic material obtained from 45 kg tissue was purified about 200 000 fold, yielding 150-200 nmoles pure NT (1).

Gel filtration on Sephadex G-25 column gave an active material with a molecular weight of 1,600 in a good agreement with the amino acid composition, indicating the presence of 13 amino acids in the peptide (1). Later, NT was isolated using

radio-immunoassay (RIA) (5) when synthetic NT became available (6). Using immunogens prepared by coupling NT through its tyroglobulin and succinylated hemocyanin several antisera toward NT have been obtained. Recently a new antiserum (TG-1) specific for the NH₂-terminal region of NT was generated. While native NT obtained from hypothalamic and intestinal extracts (7, 8) gave equal measurements with the different antisera, material extracted from other tissues gave disparate results (7). All NT-like substances reacted more strongly with COOH-terminal-directed antisera than with NH₂-terminal specific antisera and this suggested that they share in common with NT, its biologically active COOH-terminal region.

STRUCTURE

The structure of NT was deduced from sequence studies on the intact peptide and on its tryptic, chymotryptic and papain-generated fragments (2). It was suggested that NT differed slightly from this structure in situ while the pyrrolidine carboxylic acid (<Glu¹) and glutamic acid (Glu⁴) residues may have arisen by acid-catalyzed alterations of glutamine residues originally at these positions (9).

The amino acid sequence of NT doesn't resemble the other known peptides or proteins but aligned from their NH₂-terminal, vasopressin and luteinizing hormone-releasing hormone (LRH) are found to be more closely related to NT (Table 1). These sequence similarities might reflect a

Table 1. Comparison of the primary structure of neurotensin with other biologically active peptides

Neurotensin	<Glu - Leu - Tyr - Glu - Asn - Lys - Pro - Arg - Arg - Pro - Tyr - Ile - Leu - OH
Xenopsin	<Glu - Gly - Lys - Arg - Pro - Trp - Ile - Leu - OH
Vasopressin	H - Cys - Tyr - Phe - Gln - Asn - Cys - Pro - ----- - Arg - Gly - NH ₂
LRH	<Glu - His - Trp - Ser - Tyr - ---- - Gly - Leu - Arg - Pro - Gly - NH ₂
Oxytocin	H - Cys - Tyr - Ile - Gln - Asn - Cys - Pro - Leu - Gly - NH ₂
TRH	<Glu - His - Pro - NH ₂
Substance P	H - Arg - Pro - Lys - Pro - Gln - Gln - Phe - Phe - Gly - Leu - Met - NH ₂
Bradykinin	H - Arg - Pro - Pro - Gly - Phe - Ser - Pro - Phe - Arg - OH
Angiotensin I	H - Asp - Arg - Val - Tyr - Val - His - Pro - Phe - His - Leu - OH

Identical residues

acceptable codon substitutions

common origin for those three peptides (2). Only one non-mammalian peptide, xenopsin shows a remarkable resemblance to NT and exhibits a number of the biologic properties of NT (10).

NT was chemically synthesized by using the Merrifield Solid Phase procedure in 1975 and extensive studies showed that the synthetic product was chemically and biologically indistinguishable from the native material (5). Studies with synthetic NT indicated that most of the biological actions of NT resides within its COOH-terminal five to six residues (Table 2).

A proposed model of a NT-receptor interaction is shown in Figure 1. The structure-activity data was found to be consistent with this model and it provided overwhelming support for the importance of the COOH-terminal residues as specific

binding site and biological action. COOH-terminal sequences consisting five and more amino acids induced hypertension and hyperglycemia, increased vascular permeability and contracted the isolated guinea pig ileum while NH_2 -terminal sequences, as large as NT (1-10) decapeptide, were ineffective. Although the NT (9-13) pentapeptide displayed reduced potency, it possessed full intrinsic biological activity in four test systems. This indicates that whole structure is necessary for complete receptor activation. Arginin⁸ has an important role in binding since the NT (8-13) hexapeptide showed 30-50% biologic potency relative to NT. Dependence of receptor affinity on the presence of Arg⁸, Arg⁹ and a free carboxyl group of Leu¹³ indicates the involvement of strong ionic interactions between these groups and charged areas on the receptor (Fig 1) (9).

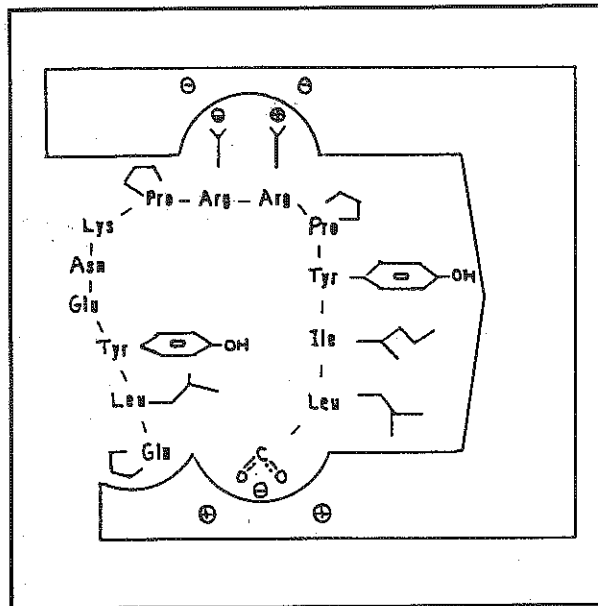


Figure 1. Proposed model for neurotensin - receptor interaction

Table 2. Relative biologic potencies of neurotensin fragments and analogs

Peptide	Percent of neurotensin activity	
	Hyperglycemia	Hypotension
<Glu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu-OH	100	100
<Glu-Lcu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile--OH	<0.5	<0.5
<Glu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-OH	<0.2	<0.2
H-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu-OH	100	80
H-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu-OH	25	20
H-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu-OH	20	10
H-Arg-Arg-Pro-Tyr-Ile-Leu-OH	55	60
H-Arg-Pro-Tyr-Ile-Leu-OH	1.0	0.5
H-Pro-Tyr-Ile-Leu-OH	<0.1	<0.1
H-Tyr-Ile-Leu-OH	<0.1	—
H-Ile-Leu-OH	<0.1	—
<Glu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu-NH ₂	<0.1	<0.1
H-Arg-Arg-Pro-Tyr-Ile-Gly-OH	<0.1	<0.1
H-Arg-Arg-Pro-Tyr-Ile-Asp-OH	<0.1	<0.1
(Xenopsin) <Glu-Gly-Lys-Arg-Pro-Trp-Ile-Leu-OH	20	20

A large number of NT analogs have been examined for their ability to interact with brain (11) or mast cell (12) receptors and for their various biological effects (13, 14). The COOH-terminal region is thought to be responsible for both binding and activity (15).

LOCALIZATION AND DISTRIBUTION

Central nervous system

Radioimmunological, immunohistochemical and radioautographic studies have shown that NT is widely distributed throughout the central nervous system of various mammalian species (16, 17). High ^{125}I -NT binding densities were observed in the bed nucleus of the stria terminalis, Celleja island, claustrum, olfactory tubercle and central nucleus of the amygdala with the exception of the central grey of the mesencephalon and interpeduncular nucleus. The remaining mesencephalon, medulla, pons, cerebellum and pineal gland have relatively low NT content (13-15).

NT has been shown to be present in the adrenal medullary cells of rat cow and cat mainly associated with chromaffin granules containing noradrenaline. It is secreted by splanchnic nerve stimulation simultaneously with catecholamines, enkephalins and somatostatin, following cholinergic receptor activation or depolarization by potassium (16).

The nerves longitudinally oriented network around large cerebral arteries and ap-

peared to be involved in modulating cerebrovascular tone in hypertension also have been shown to contain large amount of NT (17).

Gastrointestinal tract

NT is present in high concentrations in the mammalian gut, especially in endocrine cells of mucosa. Autoradiograms of ^{125}I -neurotensin bound to cross sections of the proximal and distal jejunum showed that the highest densities of silver grains were associated with the internal submucosal ganglia, external submucosal plexus and myenteric ganglia. A limited population of NT immunoreactive cells were found within the mucosal epithelium (18). Apical pole of cells containing NT in gut extends toward the intestinal lumen and these cells distributed along the villi with a characteristic morphology (19). Holzer et al. (20) reported that NT-like immunoreactivity corresponds to intact NT which was essentially distributed in the distal part of small intestine. Orci et al. (21) showed that this peptide was located mainly in discrete endocrine N cells of the mucosa. Specific NT receptor sites have been characterized in guinea pig and dog ileum smooth muscle tissues (18, 22) in which they are thought to modulate the contractility of gastrointestinal smooth muscle (23).

Plasma

Neurotensin has been detected in the range of 20 to 80 fmol/ml in extracts of

rat, bovine, dog, pig, rabbit, chicken and human plasma (7, 24, 25). Although the form of NT-like material found in plasma is not clearly known, some recent studies indicated that some of the immunoreactive materials obtained from plasma appeared to be authentic NT with the level of about 15-25 fmol/ml which is quite similar to the one identified in extracts of stomach mucosa (7).

NEUROTENSIN BINDING SITES

The existence of specific binding sites for NT has been established in the central nervous system (11, 14, 26-28). Strong hybridization signals for NT receptor mRNA were observed over neurons in the diagonal band, medial septal nucleus, nucleus basalis, magnocellularis, suprachiasmatic nucleus, substantia nigra, central tegmental area (29), hyperstriatum ventrale, archistriatum, neostriatum intermedium (30). Another study indicated that NT receptors were mainly localized in somatic sensory and motor regions of the human spinal cord suggesting that NT might play a role in modulating sensory-motor functions in human spinal cord (31).

The neurotensin receptor protein, solubilized from bovine cerebral cortex membrane exhibited saturable and specific binding of 3-11 NT with an affinity of $K_d = 5.5$ nM. It was shown that this receptor was consisted of a single polypeptide chain with a Mr of 50 000 and had intramolecular disulfide bonds (32).

In another study a soluble type of NT receptor with a molecular weight of 100

000 has been shown to contain a single binding site with a K_d of 0.36 nM and a B_m of 63 fmol/mg under both native and denaturing conditions (33). Nakagava et al. (34) examined NT binding on membrane fraction from neuroblastoma X glioma NG 108-15 hybrid cells and showed that there has been a single class of NT binding sites with a K_d of 0.86 nM and a B_{max} of 250 fmol/mg. Schotte et al. (35) demonstrated the presence of two binding sites for NT in rat brain: Site 1 was highly specific for NT and was homogeneously distributed throughout all rat brain areas while site 2 had a lower affinity for NT and its binding was markedly different in separate brain regions. It is suggested that site 1 has to be considered as an acceptor for [3H] neurotensin. Structural studies of NT binding sites in membranes of rat, rabbit and mouse brains were found in good correlation with these data (26, 36).

High NT binding densities were observed in many areas of human basal forebrain, where acetylcholinesterase is widely distributed suggesting that endogenous NT might directly influence forebrain cholinergic function in the central nervous system (37, 38). NT-receptor sites were found to be associated with presynaptic nigrostriatal dopaminergic terminals in the caudate-putamen and with glial cells in brain (39, 40). A dopaminergic control of striatal and postsynaptic NT systems were shown by several authors (40-42).

It has been previously shown that NT receptor occupancy in the adenocarcinoma HT29 cell line leads to inositol phosphate formation (43). It was suggested that NT

stimulated inositol phospholipid metabolism and caused calcium mobilization. It was also demonstrated that GTP binding-proteins were involved in this process (44-47). It was reported that NT was able to regulate intracellular Ca^{2+} levels by using inositol triphosphate as a second messenger (48-51). It was concluded that NT attenuated cAMP production by exerting an inhibitory effect on adenylate cyclase through an interaction of the peptide receptors with the regulatory GTP-binding protein N_i (52) and stimulated intracellular cGMP formation (53, 54).

A functional cDNA clone for the NT receptor recently was isolated. This receptor consisted of 424 amino acids with seven putative transmembrane domains, shows selective, high affinity binding to NT. It was concluded that it belonged to the family of G-protein-coupled-receptors and mediated its neuronal and peripheral actions by effecting the G-protein-associated second messenger system (55).

NT receptors were also found in smooth muscle of small intestine (23, 56), gastric fundus (57, 58), duodenum, ileum (59), and jejunum (18). It was concluded that NT receptors in GI system were somehow linked to ionic channels, especially a voltage-dependent calcium channel (58, 60).

BIOLOGICAL ACTIONS

Gastrointestinal effects

It was previously reported that NT contracted guinea pig ileum, relaxed rat duodenum (1,61) and effected the motility of an-

tral or fundic pouches (62). Kitabgi et al. (63) demonstrated a biphasic effect of NT application of the peptide induced rapid relaxation followed by contraction. Atropine partially inhibited NT-induced contractions of the ileum suggesting that NT might act directly on smooth muscle to relax ileum but indirectly to induce contractions. Recently, it was reported that picomolar concentrations of the NT caused concentration-dependent contractions of rat fundus. This potency was not modified with atropine and the contraction was followed by tachyphylaxis suggesting that NT activated a specific excitatory receptor located on smooth muscles of the rat fundus (58). There has been some reports indicating a central action of NT on gastric acid secretion. This inhibitory effect was thought to be occurred within nucleus accumbens and be mediated by central nervous system alpha-adrenergic receptor activation (64).

Neurologic effects

NT is distributed widely throughout the central and the peripheral nervous system suggesting a role as neurotransmitter or neuromodulator. NT has been shown to be present in the adrenal medullary cells of many mammals associated with chromaffin granules containing noradrenaline and is secreted by splanchnic nerve stimulation simultaneously within catecholamines, enkephalines and somatostatine following cholinergic receptor activation or depolarization by potassium. The presence of NT in the preganglionic nerves suggests a possible role for NT as a neuro-

transmitter (16). The relative potencies of various fragments of NT in competing for binding to brain membranes parallel in a general way their relative pharmacologic activities on peripheral tissues, but it is unclear whether the sites mediating pharmacological effects of NT in the periphery are identical with those in the central nervous system (11).

It is reported that NT directly influences the activity of magnocellular cholinergic neurons in the human forebrain (14, 37), and may be involved in the pathophysiology of demencing disorders such as Alzheimer's disease, in which these neurons have been shown to be effected (66). Significant decreases of NT receptors were found in brains of patients with Parkinson's disease (67-69). A relative NT deficiency was shown in a subgroup of psychotic patients and significant increases in NT were found after neuroleptic treatment (42, 70, 71).

The possible involvement of NT in the regulation of respiratory drive has been tested on respiratory related neurons. A bilateral apneustic pattern was induced on the phrenic nerve activities by NT injection (0.23-0.54 pmol/s) and it was suggested that NT regulates respiratory rhythmogenesis by increasing the respiratory duration (72).

A significant fall in body temperature of animals by intracisternal injection of NT was reported earlier (73). The COOH-terminal of NT was found necessary for the production of hypothermia (74), since carboxyl fragment of the molecule was shown as essential both for binding to

brain membranes and for biologic activity (27). Nemeroff et al. (75) found that central injection of NT increased sleeping time in mice treated with pentobarbital. Recently, when ethanol and NT were administered in combination, sedative and hyperthermic effects of these two substances were found to be potentiated (76). Kalivas (77) reported that NT could act in the ventral tegmental area and ventromedial mesencephalon to produce both a decrease in colonic temperature and an increase in spontaneous motor activity.

Endocrine effects

A single i.v. injection of NT induces dose-related hyperglycemia in animals (78-80). Carraway et al. (81) observed that the increase in plasma glucose was associated with dose-dependent increase in hepatic glycogen phosphorylase activity and reduction in liver glycogen content. They suggested that NT injection raised blood glucose by increasing hepatic gluconeogenesis. Wolfe et al. (79) observed a rise in plasma glucose levels and glucose production in glycogen-depleted animals following NT injection and concluded that this reflected an enhanced rate of gluconeogenesis. High doses of NT injection also reduces plasma insulin levels and increases circulating glucagon levels (78, 82, 83). The addition of NT to isolated pancreas preparations suppressed insulin secretion and stimulated glucagon release into the medium (84). However, a dual effect of NT on pancreatic hormone secretion was observed: at low concentrations, NT stimulated insulin, glucagon and somatostatin

release whereas at high concentrations it suppressed the release of these hormones (85). NT was shown to modulate the secretion of pancreatic hormones (83, 86).

The multiple effects of NT on gluco-regulation may be mediated by histamine or catecholamines. Diphenhydramine, an H_1 receptor antagonist, blocked the hyperglycemia and hyperglucagonemia caused by the injection of NT or histamine while H_2 receptor blockade did not effect the response. NT injection also was found to be associated with a rise in plasma insulin levels (83, 87).

NT effects the release of several pituitary hormones. I.v. injection of NT caused increases at circulatory levels of prolactin, growth hormone, corticosteroids and ovarian steroids (88-92).

Vascular Effects

I.v injection of NT (0.16-1.0 mg/kg) caused a dose -related fall in arterial blood pressure in rats and acute tachyphylaxis accompanied the first administration of the peptide (1). Rapid vasodilatation of small blood vessels with NT injection has been reported previously and this response has been correlated with an increase in regional blood flow to the intestine (93). It was suggested that NT injection caused a rapid increase in vascular permeability to protein, an associated increase in hematocrite and a reversible cyanosis lasting 5-10 minutes (1). Induction of hypotension by NT was found to be mediated by histamine (16).

REFERENCES

1. Carraway, R., Leeman, S.E., "The Isolation of a New Hypotensive Peptide. Neurotensin, from Bovine Hypothalami", *J. Biol. Chem.*, 248 (19), 6854-6851, 1973.
2. Carraway, R., Leeman, S.E., "The Amino Acid Sequence of a New Hypothalamic Peptide. Neurotensin", *J. Biol. Chem.*, 250 (5), 1907-1911, 1975.
3. Carraway, R., Kitabgi, P., "The Amino Acid Sequence of Radioimmunoassayable Neurotensin from Bovine Intestine", *J. Biol. Chem.*, 253, 7996-7998, 1978.
4. Hammer, R.A., Leeman, S.E., "Isolation of Human Intestinal Neurotensin", *J. Biol. Chem.*, 255, 2476-2480, 1980.
5. Carraway, R., Leeman, S.E., "Radioimmunoassay for Neurotensin, a Hypothalamic Peptide", *J. Biol. Chem.*, 251 (22), 7035-7044, 1976.
6. Carraway, R., Leeman, S.E. "The Synthesis of Neurotensin", *J. Biol. Chem.*, 250 (5), 1912 - 1918, 1975.
7. Carraway, R., Leeman, S.E., "Characterization of Radioimmunoassayable Neurotensin in the Rats. Its Differential Distribution in the central Nervous System, Small Intestine and Stomach", *J. Biol. Chem.*, 251 (22), 7025-7052, 1976.

8. Kitabgi, P., Carraway, R., "Isolation of a Tridecapeptide from Bovine Intestine Tissue and its Partial Characterization as Neurotensin", *J. Biol. Chem.*, 251 (22), 7053-7058, 1976.
9. Chang, D., Humphries, J., "Synthesis and Activities of Neurotensin and its Amidated Analogs and Possible Natural Occurance of (Gln⁴) - Neurotensin", *Proc. Natl. Acad. Sci. USA.*, 73 (11), 3833-3837, 1976.
10. Ishida, T., Kawamura, K., "Comparison Studies of Neurotensin and Xenopsin upon Pancreatic Secretion in the Dog", *Metabolism* (Suppl 1), 25 (11), 1467-1468, 1976.
11. Uhl, G.R., Bennett, J.P., "Neurotensin, a Central Nervous System Peptide; Apparent Receptor Binding in Brain Membranes", *Brain Res.*, 130, 299-313, 1977.
12. Lazarus, L.H., Perrin, M.H., "Verification of Both the Sequence and Conformational Specificity of Neurotensin in Binding to Mast Cells", *Biochem. Biophys. Res. Commun.*, 76, 1079-1085, 1977.
13. Uhl, G.R., Kuhar, M.J., Synder, S.H., "Neurotensin: Immunohistochemical Localization in Rat Central Nervous System", *Proc. Natl. Acad. Sci.*, 74 (9), 4059-4063, 1977.
14. Szigethy, E., Quirion, R., Beaudet, A., "Distribution of ¹²⁵I-Neurotensin Binding Sites in Human Forebrain: Comparison with the Localization of Acetylcholinesterase", *J. Comp. Neurol.*, 297 (4), 487-498, 1990.
15. Kobayashi, R.M., Brown, M. Vale, W., "Regional Distribution of Neurotensin and Somatostatin in rat Brain", *Brain Res.*, 126, 584-588, 1977.
16. Bommer, M., Herz, A., "Neurotensin Affects Metabolism of Opioid Peptides, Catecholamines and Inositol Phospholipids in Bovine Chromaffin cells", *Life Sci.*, 44, 327-335, 1989.
17. Young, W.S., Kuhar, M.J., "Neurotensin Receptor Localization by Light Microscopic Autoradiography in Rat Brain", *Brain Res.*, 206, 273-285, 1981.
18. Seybold, V.S., Treder, B.G., Aannonsen, L.M., "Neurotensin Binding Sites in Porcine Jejunum: Biochemical Characterization and Intramural Localization", *Synapse*, 6 (1), 81-90, 1990.
19. Sundler, F., Aluments, J., Hakanson, R., "Ultrastructure of the Gut Neurotensin Cell", *Histochemistry*, 53, 25-34, 1977.
20. Holzer, P., Bucsics, A., Saria, A., "A study of Concentrations of Substance P and Neurotensin in the Gastrointestinal Tract of Various Mammals", *Neuroscience*, 7, 2917-2919, 1982.

21. Orci, L., Baetens, O., Rufener, C., Brown, M., "Evidence for Immunoreactive Neurotensin in Dog, Intestinal Mucosa", *Life Sci.*, 19, 559-562, 1976.
22. Mitra, S.P., Muraki, T., Brown, D.R., "Canine Neurotensin, Neurotensin 6-13 and Neuromedin N: Primary Structures and Receptor Activity", *Regul. Pept.*, 28 (1), 11-22, 1990.
23. Ahmad, S., Daniel, E.E., "Neurotensin Receptors on Circular Smooth Muscle of Canine Small Intestine: Role of Disulfide Bridges", *Biochem. Biophys. Res. Commun.*, 165 (1), 422-428, 1989.
24. Carraway, R., Hammer, R.A., Leeman, S.E., "Neurotensin in Plasma: Immunochemical and Chromatographic Character of Acid/Acetone Soluble Material", *Endocrinology*, 107, 400-407, 1980.
25. Shulkes, A., Click, P., Wang, H., "A Radioimmunoassay for Neurotensin in Human Plasma", *Clin. Chim. Acta*, 125, 49-55, 1982.
26. Mazella, J., Chabry, J., "Purification of the Neurotensin Receptor from Mouse Brain by Affinity Chromatography", *J. Biol. Chem.*, 264, 5559-5563, 1989.
27. Kitabgi, P., Carraway, R., "Neurotensin: Specific Binding to Synaptic Membranes from Rat Brain", *Proc. Natl. Acad. Sci. USA*, 74 (5), 1846-1850, 1977.
28. Lazarus, L.H., Brown, M., Perrin, M.H., "Distribution, Localization and Characteristics of Neurotensin Binding Sites in the Rat Brain", *Neuropharmacology*, 16, 625-629, 1977.
29. Elde, R., Schalling, M., Ceccatelli, S., "Localization of Neuropeptide Receptor mRNA in Rat Brain: initial Observations Using Probes for Neurotensin and Substance P receptor", *Neurosci. Lett.*, 120 (1), 134-138, 1990.
30. Brauth, S.E., Kitt, C.A., "Neurotensin Binding Sites in the Forebrain and Midbrain of the Pigeon", *J. Comp. Neurol.*, 253 (3), 358-373, 1986.
31. Awad, E.W., Nassar, C.F., Tabbara, M.S., "Characteristics and Displaceability of Neurotensin Binding Sites in the Rat Cerebral Cortex and Corpus Striatum", *Gen. Pharm.*, 20 (6), 725-729, 1989.
32. Mills, A., Demoliou-Mason, C.D., Bernard, E.A., "Purification of the Neurotensin Receptor from Bovine Brain", *J. Biol. Chem.*, 263 (1), 13-16, 1988.
33. Mazella, J., Chabry, J., "Solubilization and Characterization of Active Neurotensin Receptors from Mouse Brain", *J. Biol. Chem.*, 263, 144-149, 1988.

34. Nakagawwa, Y., Higashida, H., "A Single Class of Neurotensin Receptors with High Affinity in Neuroblastoma X Glioma NG 108-15 Hybrid Cells that Mediate Facilitation of Synaptic Transmission", *J. Neurosci.*, 4 (6), 1653-1661, 1984.
35. Schotte, A., Leysen, J.E.: Evidence for Displace Non-Specific [3H]-neurotensin Binding Site in Rat Brain", *Naunyn. Schmiedebergs. Arch. Pharm.*, 33 (4), 400-405, 1986.
36. Mazella, J., Kitabgi, P., "Molecular Properties of Neurotensin Receptors in Rat Brain", *J. Biol. Chem.*, 260, 508-514, 1985.
37. Szigethy, E., Beaudet, A., "Selective Association of Neurotensin Receptors with Cholinergic Neurons in the Rat Basal Forebrain", *Neurosci. Lett.*, 83 (1-2), 47-52, 1987.
38. Szigethy, E., Leonard, K., "Ultrastructural Localization of ¹²⁵I Neurotensin Binding Sites to Cholinergic Neurons of the Rat Nucleus Basalis Magnocellularis", *Neuroscience*, 36 (2), 377-391, 1990.
39. Schotte, A., Rostene, W., "Different Subcellular Localization of Neurotensin-receptor and Neurotensin-acceptor Sites in the Rat Brain Dopaminergic System", *J. Neurochem.*, 50 (4), 1026-1031, 1988.
40. Qirion, R., Welner, S., Gauthier, S., "Neurotensin Receptor Binding Sites in Monkey and Human Brain: Autoradiographic Distribution and Effects of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine Treatment", *Synapse*, 1 (6), 559-566, 1987.
41. Masuo, Y., Pelaprat D., "Regulation of Neurotensin-containing Neurons in the Rat Striatum and Substantia Nigra. Effects of Unilateral Nigral Lesion with 6-hydroxydopamin on Neurotensin Content and Its Binding Site Density", *Brain. Res.*, 510 (2), 203-210, 1990.
42. Herve, D., Tassin, J.P., "Dopaminergic Control of ¹²⁵I-labeled Neurotensin Binding Site Density in Corticolimbic Structures of the Rat Brain", *Proc. Natl. Acad. Sci. USA*, 83 (16), 6023-6207, 1986.
43. Bozou, J.C., Rochet, N., Magnaldo, I., "Neurotensin Stimulates Inositol-Triphosphate-Mediated Calcium Mobilization but not Protein kinase C Activation in HT29 Cells. Involvement of a G-protein", *Biochem. J.*, 264 (3), 871-878, 1986.
44. Snider, R.M., Furray, C., Phenig, M., "Neurotensin Stimulates Inositol Phospholipid Metabolism and Calcium Mobilization in Murine Neuroblastoma Clone N1E-115", *J. Neurochem.*, 47 (4), 1214-1218, 1986.
45. Amar, S., Kitabgi, P., "Stimulation of Inositol Phosphate Produc-

- tion by Neurotensin in Neuroblastoma N1E-115 Cells: Implications of GTP-Binding Protein and Relationship with the Cyclic GMP Response", *Neurochem.*, 49, 999-1006, 1987.
46. Goedert, M., Pinnock, R.D., "Neurotensin Stimulates Inositol Phospholipid Hydrolysis in Rat Brain Slices", *Brain. Res.*, 323, 193-197, 1987.
47. Amar, S. Kitabgi, P., "Activation of Phosphatidylinositol Turnover by Neurotensin Receptors in the Human Clonogenic Adenocarcinoma Cell Line", *FEBS Lett.*, 201 (1), 31-36, 1986.
48. Donoso, M.V., "Involvement of Calcium Channels in the Contractile Activity of Neurotensin but not Acetylcholine: Studies with Calcium Channel Blockers and Bay K 8644 on the Rat Fundus", *Br. J. Pharmacol.*, 88 (4), 837-846, 1986.
49. Sato, M., Shiosaka, S., Tohyama, M., "Neurotensin and Neuromedin N Elevate the Cytosolic Calcium Concentration via Transiently Appearing Neurotensin Binding Sites in Cultured Rat Cortex Cells", *Brain. Res. Dev. Brain. Res.*, 58 (1), 97-103, 1991.
50. Wu, H., Franklin, C.C., "Regulation of Calcium Activated Potassium Efflux by Neurotensin and Other Agents in HT209 Cells", *Am. J. Phys.*, 260 (1 pt), C35-41, 1991.
51. Turner, J.T., Kracke, J.M.R., "Regulation of the Neurotensin Receptor and Intracellular Calcium Mobilization in HT29 cells", *J. Pharmacol. Exp. Ther.*, 253 (3), 1049-1056, 1990.
52. Bozoğ̃u, J.C., Amar, S., Vincent J.P., "Neurotensin-mediated Inhibition of Cyclic AMP Formation in Neuroblastoma N1E-115 Cells: Involvement of the Inhibitory GTP-Binding Component of Adenylate Cyclase", *Mol. Pharmacol.*, 29 (5), 489-496, 1986.
53. Gilbert, J.A., Moses, C.J.: Neurotensin and Its Analogs: Correlation of Specific Binding with Stimulation of Cyclic GMP Formation in Neuroblastoma Clone N1E-115", *Biochem. Pharmacol.*, 35 (3), 391-397, 1986.
54. Amar, S., Mazella, J., Checler, F., "Regulation of Cyclic GMP Levels by Neurotensin in Neuroblastoma Clone N1E-115", *Biochem. Biophys. Res. Commun.*, 129 (1), 117-125, 1985.
55. Tanaka, K., Masu, M., "Structure and Functional Expressions of the Closed Rat Neurotensin Receptor", *Neuron*, 4 (6), 847-854, 1990.
56. Ahmad, S., Berezin, I., "Neurotensin Receptors in Canine Intestinal Smooth Muscle: Preparation of Plasma Membranes and Characterization of (Tyr³-125I) Labelled Neurotensin Binding", *Biochim.*

- Biophys. Acta*, 986 (2), 224-228, 1987.
57. Mazella, J., Kwan, C.Y., "Covalent Labeling of Neurotensin Receptors in Rat Gastric Fundus Plasma Membranes", *Peptides* 6 (6), 1137-1141, 1985.
 58. Huidobro, T.J.P., Kullak, A., "Excitatory Neurotensin Receptors on the Smooth Muscle of the Rat Fundus: Possible Implications in Gastric Motility", *Br. J. Pharm.*, 84 (4), 897-910, 1985.
 59. Donoso, M.V., Huidobro, T.J.P., "Gastrointestinal Neurotensin Receptors: Lack of Modulation by Thyrotropin Releasing Hormone", *J. Pharm. Pharmacol.*, 37 (6), 425-428, 1985.
 60. Katsoulis, S., Conlon, J.M., "Neurotensin and Prostaglandin Interaction in Smooth Muscle of the Guinea Pig Stomach", *Eur. J. Pharmacol.*, 158 (3), 251-256, 1988.
 61. Rokeaus, A., Burcher, E., "Actions of Neurotensin and (Gln⁴)-neurotensin on Isolated Tissues". *Acta Pharmacol. Toxicol.*, 41, 141-147, 1977.
 62. Andersson, S., Rosell, S., "Inhibition of Gastric and Intestinal Motor Activity in Dogs by (Gln⁴)-Neurotensin", *Acta Physiol. Scand.*, 100, 231-235, 1977.
 63. Kitabgi, P., Freychet, P., "Effect of Neurotensin on Isolated Intestinal Smooth Muscles", *Eur. J. Pharmacol.*, 50, 349-357, 1978.
 64. Vhang, L. Xing, L.P., "Central Neurotensin Inhibits Gastric Acid Secretion: An Adrenergic Mechanism in Rats", *Gastroenterology*, 97 (5), 1130-1134, 1989.
 65. Huidobro, T.J.P., Zhu, Y.X., "Neurotensin Receptors on the Ileum of the Guinea Pig: Evidence for the Coexistence of Inhibitory and Excitatory Receptors", *Eur. J. Pharmacol.*, 102 (2), 237-250, 1984.
 66. Jansen, K.L., Faull, R.L., "Alzheimer's Disease: Changes in Hippocampal N-methyl-D-Aspartate, Quisqualate, Neurotensin, Adenosin, Benzodiazepine, Serotonin and Opioid Receptors. An Autoradiographic Study", *Neuroscience*, 39 (3), 613-627, 1990.
 67. Chinaglia, G., Probst, A., "Neurotensin Receptors in Parkinson's Disease and Progressive Supranuclear Palsy: An Autoradiographic Study in Basal Ganglia", *Neuroscience*, 39 (2), 351-360, 1990.
 68. Uhl, G.R., Hackney, G.O., "Parkinson's Disease: Nigral Receptor Changes Support Peptidergic Role in Nigrostriatal Modulation", *Ann. Neurol.*, 20 (2), 194-203, 1986.
 69. Sadoul, J.L., Checla, F., "Loss of High Affinity Neurotensin Receptors in Substantia Nigra from Par-

- kinsonian Subjects". *Biochem. Biophys., Res. Commun.*, 125 (1), 395-404, 1984.
70. Garver, D.L., Bisette, G., Yao, J.K., "Relation of CSF Neurotensin Concentrations to Symptoms and Drug Response of Psychotic Patients", *Am. J. Psychiatry.*, 148 (4), 484-488, 1991.
71. Giardino, L., Calza, L., "DA2/NT Receptor Balance in the Mesostriatal and Mesolimbocortical Systems After Chronic Treatment with Typical and Atypical Neuroleptic Drugs", *Brain Res.*, 532 (1-2), 140-145, 1990.
72. Morin-Surun, M.P., "The Excitation by Neurotensin of Nucleus Tractus Solitarius Neurons Induced Apneustic Breathing", *Brain Res.*, 106-113, 1986.
73. Bisette, G., Nemeroff, C.B., "Hypothermia and Intolerance to Cold Induced by Intracisternal Administration of the Hypothalamic Peptide Neurotensin", *Nature*, 262, 607-609, 1976.
74. Loosen, P.T., Nemeroff, C.B., Bisette, G., "Neurotensin-induced Hypothermia in the Rat: Structure Activity Studies", *Neuropharmacology*, 17, 109-113, 1978.
75. Nemeroff, C.B., Bisette, G., Prange, A.J.Jr., "Neurotensin: Central Nervous System Effects of a Hypothalamic Peptide", *Brain Res.*, 128, 485-496, 1977.
76. Smith, T.L., "The Effects of Acute Exposure to Ethanol on Neurotensin and Guanine Nucleotide-Stimulation of Phospholipase C Activity in intact N1E-115 Neuroblastoma Cells", *Life Sci.*, 47 (20), PL115-119, 1990.
77. Kalivas, P.W., "Neurotensin in the Ventromedial Mesencephalon of the Ratanatomical and Functional Considerations", *J. Comp. Neurol.*, 226 (4), 495-507, 1984.
78. Brown, M., Vale, W., "Effect of Neurotensin and Substance P on Plasma Insulin, Glucagon and Glucose Levels", *Endocrinology*, 98, 819-821, 1976.
79. Wolfe, R.R., Allsop, J.R., Burke, J.F., "Increased Glucose Production Following Neurotensin Administration", *Life Sci.*, 22, 1043-1048, 1978.
80. Kaneto, A., Kaneko, T., "Effect of Substance P and Neurotensin Infused Intrapancreatically on Glucagon and Insulin Secretion", *Endocrinology*, 102, 393-401, 1978.
81. Carraway, R., Demers, L.M., Lee-man, S.E., "Hyperglycemic effect of Neurotensin, a Hypothalamic Peptide", *Endocrinology*, 99, 1452-1462, 1976.
82. Rambout, J.H., Van der Grinten C.P., "Immunocytochemical Identification and Localization of Peptide Hormones in the Gastro-Entero-Pancreatic (GEP) Endo-

- crine System of the Mouse and a Stomachless fish, *Barbus Conchoni*", *Histochemistry*, 84 (4-6), 471-483, 1986.
83. Reinecke, M., "Neurotensin Immunohistochemical Localization in Central and Nervous System and in Endocrine Cells and its Functional Role as Neurotransmitter and Endocrine Hormone", *Prog. Histochem. Cytochem.*, 16 (1), 1-172, 1985.
84. Moltz, J.H., Dobbs, R.E., "Effect of Hypothalamic Factors on Insulin and Glucagon Release from the Islets of Langerhans", *Endocrinology*, 101, 196-202, 1977.
85. Dolais-Kitabgi, S., Kitabgi, P., "Effects of Neurotensin on Insulin, Glucagon and Somatostatin Release From Isolated Pancreatic Islets", *Endocrinology*, 105, 256-601, 1979.
86. Ballesta, J., Bloom, S.R., "Distribution and Localization of Regulatory Peptide", *Crit. Rev. Clin. Lab. Sci.*, 22 (3), 185-218, 1985.
87. Nagai, K., Frohman, L.A., "Hyperglycemia and Hyperglucagonemia Following Neurotensin Administration", *Life Sci.*, 19, 273-280, 1976.
88. Vijayan, E., McCann, S.M., "Effect of Intraventricular Injection of Substance P. Neurotensin and Gas-trin on Pituitary Hormone Release in Conscious, Ovariectomized rats", *Endocrinology*, 102, 271A-275A, 1978.
89. Memo, M., Castelletti, L., "Identification of Neurotensin Receptors Associated with Calcium Channels and Prolactin Release in Rat Pituitary", *J. Neurochem.*, 47 (6), 1682-1688, 1986.
90. Rivier, C., Brown, M., "Effect of Neurotensin, Substance P and Growth Hormone in the Rat", *Endocrinology*, 100, 751-754, 1977.
91. Alexander, M.J., Dobner, P.R., Miller, M.A., "Estrogen Induces Neurotensin Neuromedin N Messenger RNA in a Preoptic Nucleus Essential for the Preovulatory Surge of Luteinizing Hormone in rat", *Endocrinology*, 125 (4), 2111-2117, 1989.
92. Von Euler, G., Meister, B. "Intraventricular Injection of Neurotensin Reduces Dopamine D2 Agonist Binding in Rat Forebrain and Intermediate Lobe of the Pituitary Gland. Relationship to Serum Hormone Levels and Nerve Terminal Coexistence", *Brain Res.*, 531 (1-2), 253-262, 1990.
93. Rosell, S., Burcher, E., "Cardiovascular and Metabolic Actions of Neurotensin and (Gln⁴) Neurotensin", *Acta Physiol. Scand.*, 98, 484-491, 1976.