

Search for Antifungal Compounds from Some *Verbascum* Species Growing in Turkey

İ. İrem TATLI*^o, Zeliha Ş. AKDEMİR*, Erdal BEDİR**,*^{***}, İkhlas A. KHAN**

Search for Antifungal Compounds from Some *Verbascum* Species Growing in Turkey

Summary : Turkish medicinal plants provide a rich source of biologically-active natural products. With the use of simple bioassays, it is possible to evaluate the bioactivity of the natural compounds. Antifungal screening of 16 compounds in a matrix format from two *Verbascum* species growing in Turkey was conducted directly on thin-layer chromatographic (TLC) plates sprayed with a spore suspension. Compounds possessing strong antifungal activity produced a clear zone of inhibition bounded by a sharp margin regardless of the size of the inhibitory zone. Ilwensisaponin A and C from *Verbascum pterocalycinum* var. *mutense* Hub.-Mor. were found to be active. Bioautographic assay indicated that the saponins appeared to be the most effective against *Colletotrichum acutatum*, *C. fragariae* and *C. gloeosporioides*

Keywords: Medicinal plants, *Verbascum lasianthum*, *Verbascum pterocalycinum* var. *mutense*, Scrophulariaceae, Bioactive Compounds, Saponins, Ilwensisaponin A, Ilwensisaponin C, Antifungal Activity

Received : 28.01.2004

Revised : 18.03.2004

Accepted : 30.03.2004

Türkiye'de Yetişen Bazı *Verbascum* Türlerinden Elde Edilen Antifungal Bileşiklerin Araştırılması

Özet : Türkiye'de yetişen tıbbi bitkiler, biyolojik olarak aktif doğal ürünlerce zengin bir kaynak oluşturmaktadırlar. Basit biyolojik tarama metodları kullanarak, doğal bileşiklerin biyolojik aktivitelerini belirlemek mümkündür. Türkiye'e yetişen iki *Verbascum* türünden elde edilen 16 bileşiğin antifungal aktivite taraması, spor süspansiyonu püskürtülmüş ince tabaka kromatografisi (İTK) plaklarına direkt olarak uygulanmaları ile yapılmıştır. Kuvvetli antifungal aktiviteye sahip bileşikler, inhibisyon zonlarının boyutları gözlemlenmesinin keskin sınırları olan temiz bir zon vermişlerdir. *Verbascum pterocalycinum* var. *mutense*'den elde edilen ilwensisaponin A ve C aktif bulunmuştur. Biyotografik metot ile, saponinlerin *Colletotrichum acutatum*, *C. fragariae* ve *C. gloeosporioides*'e karşı en fazla etkiyi gösterdiği görülmüştür

Anahtar kelimeler: Tıbbi Bitkiler, *Verbascum lasianthum*, *Verbascum pterocalycinum* var. *mutense*, Scrophulariaceae, Biyoaktif Bileşikler, Saponinler, Ilwensisaponin A, Ilwensisaponin C, Antifungal Aktivite

INTRODUCTION

In many developing countries, plants constitute the main medicines used in health care practice¹. Most of the plants are regularly used as traditional medicine for the treatment of various diseases in Turkey^{2,3}. Consequently, it is important to evaluate the utilized plants and remedies in order to gain more knowledge about the efficacy of this approach. The choice of plant material for such studies can be guided by consulting traditional healers, herbalists or the documented information¹.

Verbascum, commonly known as "mullein", is a widespread genus of the family Scrophulariaceae, and is represented by 228 species (185 of which are endemic) in the flora of Turkey⁴. Some *Verbascum* species have been used as piscicide, antiseptic, astringent, demulcent, emollient, expectorant, sedative, narcotic, diuretic and antimalarial and as a treatment for tumors, inflammations, migraine, asthma and spasmodic coughs in Turkey, Europe, Asia and Northern America^{3,5}.

Some phytochemical studies on *Verbascum* species

* Hacettepe University, Faculty of Pharmacy, Department of Pharmacognosy, 06100, Sıhhiye-Ankara, TURKEY.

** National Center for Natural Products Research Institute of Pharmaceutical Sciences, University of Mississippi, University, Mississippi 38677, USA

*** Ege University, Faculty of Engineering, Department of Bioengineering, Bornova 35100, İzmir, TURKEY.

^o Corresponding author e-mail: itatli@hacettepe.edu.tr

of Turkish origin have revealed the presence of iridoids, phenylethanoids⁶, saponins and monoterpenoids⁷. Iridoids display an interesting spectrum of biological activity such as antiinflammatory⁸ and antimicrobial⁹. Likewise, phenylethanoid glycosides also show a wide range of biological activity. They are known to possess antibacterial and antifungal activities¹⁰. Saponins are also well-documented as exhibiting a variety of biological activities, including antibacterial and antifungal activity¹¹.

Owing to the continuing development of the microbial-resistance in medicine and agriculture, the discovery of new antimicrobial substances is important¹². Plant-derived constituents offer potential leads for the development of antifungal drugs effective against human pathogenic fungi¹³. In addition, the desire for safer agrochemicals with less environmental and mammalian toxicity is a major concern. Particularly desirable is the discovery of novel antimicrobial agents representing new and natural chemical classes that operate by different models of action from existing antifungal agents^{12,14,15}.

There is a growing interest in antifungal compounds as the occurrence of systematic mycoses associated with immunodeficiency diseases (such as AIDS) and the use of immunosuppressors is continually increasing¹.

Therefore, it would be interesting to study the effect of the compounds from *Verbascum* species on medically important fungi for the development of new antifungal agents for the treatment of serious fungal infections, especially in immunosuppressed and immunocompromised patients.

As a part of our continuing search for bioactive agents from natural sources, we here have report the results of the antifungal activity by thin-layer chromatography (TLC)-bioautographic assay of the compounds from *Verbascum lasianthum* and *V. pterocalycinum* var. *mutense* in different classes.

MATERIALS and METHODS

Plant Materials

The research materials were collected from the following locations.

Verbascum lasianthum Boiss. ex Benth: B1: İzmir: Urla, Üçahurlar Mevkii, August 1999.

Verbascum pterocalycinum var. *mutense* Hub.-Mor.: C4: İçel: Between Mut and Karaman, *Pinus brutia* and *Pinus nigra* fields, 930-1100 m, July 2000.

Voucher specimens were deposited in the herbarium of the Pharmacognosy Department, Faculty of Pharmacy, Hacettepe University, Ankara, Turkey (HUEF 99130 and HUEF 00184, respectively).

Microorganisms

Three fungal strains, *Colletotrichum acutatum* Simmonds, *C. fragariae* Brooks, and *C. gloeosporioides* (Penz.) Penz. & Sacc. in Penz., which are important plant pathogenic fungi, were used in the assay. The three *Colletotrichum* species were isolated from strawberry (*Fragaria x ananassa* Duchesne). Commercial fungicides vinclozolin, chlorothalonil and thiabendazole (Chem. Service, West Chester, PA) were used as validation controls in a microbioassay¹⁵.

Inoculum Preparation

Conidial suspensions were prepared according to published procedures⁸. Conidial concentrations were determined spectro photometrically^{16,17} from a standard curve, and suspensions were then adjusted with sterile distilled water to a concentration of 1.0×10^6 conidia ml⁻¹.

Antifungal Assay

Inhibition of fungal growth on chromatographic plates was evaluated by modifications of TLC bioa-

utographic assays^{18,19}. Each sample was dissolved in MeOH and commercial fungicide standards in 95% EtOH. Each test compound was applied on glass-backed silica gel plates from stock solutions to achieve a final amount of 2 µg using a disposable glass micropipette for each sample. To detect biological activity directly on the TLC plate, silica gel plates with a fluorescent indicator were sprayed with conidial suspensions of *Colletotrichum acutatum*, *C. fragariae* and *C. gloeosporioides*. Aliquots of 25-50 ml of inoculum spray solution were prepared for each test fungus with liquid potato dextrose broth (PDB) containing 12 g /500 ml (PDB), 0.1% bacto agar, and 0.1% Tween-80. Using a 50 ml chromatographic sprayer, each plate was sprayed lightly (to a damp appearance) three times with conidial suspension. Clear inhibition zones were observed against a dark-grey background after four days incubation at room temperature in a humid atmosphere.

RESULTS AND DISCUSSION

Previously isolated iridoid glucosides, 6-*O*-(α -L-rhamnopyranosyl)-catalpol (**1**), verbascoside A (**2**), pulverulentoside I (**3**), buddlejioside A₅ (**4**), aucubin (**5**), unduloside III (**6**), 8-*O*-acetylharpagide (**7**), harpagoside (**8**) and vanilloylajugol (**9**), along with two phenylethanoid glycosides, verbascoside (**10**) and poliumoside (**11**), from the roots of *Verbascum lasianthum*^{6,20}, and two saponins, ilwensisaponin A (**12**) and C (**13**), together with two iridoid glycosides, ajugol (**14**) and picoside IV (**15**) and a monoterpene glucoside, 1-(β -D-glucopyranosyl)-8-hydroxy-3,7-dimethyl-oct-2(E),6(E)-dienoate (**16**), from the flowers of *Verbascum pterocalycinum* var. *mutense*⁷ were used in this study.

Compounds **1-16** were further evaluated for *in vitro* antifungal activity on TLC-bioautographic assay^{18,19}. Active compounds appeared as clear spots against a colored background. In the test for fungicidal activity, the saponins, ilwensisaponin A and C, were found to exhibit some *in vitro* activity against *Colletotrichum acutatum*, *C. fragariae* and *C. gloeosporioides*.

The results of this survey thus showed that *Verbascum* species contain potentially bioactive saponins and that it is worth studying this antifungal assay for saponins.

CONCLUSION

To date, many saponins have been isolated and characterized from *Verbascum* species^{7,21}. In Turkey and other countries, *Verbascum* species have long been utilized for medicinal purposes, and saponins are considered to have the active principles possessing such physiological activities¹¹. Triterpene saponins are described as having interesting biological activities such as anti-inflammatory, molluscicidal and fungistatic²². Some structural features for antimicrobial activity of triterpene saponins have been documented in the literature: The strongest activities are exhibited by the monodesmosidic saponins. The availability of ester groups and an increase in the number of sugar units lead to an increase in activity²³. Maximum activity is shown by monodesmosides with four or five monosaccharides. Glycosides with monosaccharide moiety in the position C-3 of the aglycone were strongly antimycotic. One theory is that the saponins themselves are inactive and comprise only water-soluble transport forms. In the presence of cell membrane glycosidases, there is formation of the aglycone which is the active membranolytic component. It must be emphasized that other mechanisms must also be involved, since there are exceptions to the parallel between cholesterol binding and fungicidal properties¹¹.

Therefore it is proven that the antifungal activity of ilwensisaponin A and C is closely related to their structure having a monodesmosidic with tetrasaccharidic moiety at C-3.

To our knowledge this is first report of the antifungal activity of ilwensisaponin A and C.

In order to investigate the plant species for other activities related to their traditional use, further bioassays will need to be performed.

Natural product research will contribute to the long-range improvement of US agriculture by enhancing our knowledge about disease control measures and host-pathogen interactions and may provide an opportunity for the discovery of new pharmaceutical agents to treat human diseases¹⁶.

Acknowledgements

The authors thank Prof. Dr. Hayri Duman, Gazi University, Faculty of Science, Department of Botany, Etiler, Ankara, Turkey, for the authentication of the plant specimen. Antifungal assays were supported by NIH grant AI 27094. This work was also supported in part by the United States Department of Agriculture, ARS Specific Cooperative Research Agreement no. 58-6408-7-012.

REFERENCES

1. Diallo D, Marston A, Terreaux C, Toure Y, Smestad Paulsen B, Hostettmann K. Screening of Malian plants for antifungal, larvicidal, molluscicidal, antioxidant and radical scavenging activities, *Phytotherapy Research*, 15, 401-406, 2001.
2. Baytop T, Therapy with Medicinal Plants in Turkey (Past and Present), 2nd ed., Nobel Tıp Kitabevleri Ltd., İstanbul, 334- 335, 1999.
3. Sezik E, Yeşilada E, Honda G, Takaishi Y, Takeda Y, Tanaka T. Traditional medicine in Turkey X. Folk medicine in Central Anatolia, *J. Ethnopharmacol.*, 75, 95-115, 2001.
4. Huber-Morath A. *Verbascum*, Davis, PH (ed.), Flora of Turkey and the East Aegean Islands, Edinburgh University Press, 6,461-603, 1978.
5. Grieve M., A Modern Herbal, Barnes and Noble Books, New York, 564-566, 1995.
6. Akdemir ZS, Tatli II, Bedir E, Khan IA. Iridoid and phenylethanoid glycosides from *Verbascum lasianthum* Boiss., *Turk. J. Chem.*, 28, 227-234, 2004.
7. Tatli II, Akdemir ZS, Bedir E, Khan IA. Saponin, iridoid, phenylethanoid and monoterpene glycosides from *Verbascum pterocalycinum* var. *mutense* Hub.-Mor., *Turk. J. Chem.*, 28, 111-122, 2004.
8. Recio MC, Giner RM, Manez S, Rios JN. Structural considerations on the iridoids as anti-inflammatory agents, *Planta Med.*, 60, 232-234, 1994.
9. Sticher O. New Natural Products and Plant Drugs with Pharmacological, Biological or Therapeutical Activity, Springer, New York, Berlin, 1977.
10. Jimenez C, Riguera R. Phenylethanoid glycosides in plants: structure and biological activity, *Natural Product Reports*, 591-606, 1994.
11. Rao AV, Gurfinkel DM. The bioactivity of saponins: triterpenoid and steroidal glycosides, *Drug Metabolism and Drug Interactions*, Freud Publishing House Ltd., Toronto, ON, 17, 211-235, 2000.
12. McChesney JD. Biological and chemical diversity and the search for new pharmaceuticals and other bioactive natural products, Kinghorn AD, Balandrin MF (eds.), *Human Medicinal Agents from Plants*, ACS Symposium Series American Chemical Society, Washington, DC, 534, 38-47, 1993.
13. Hufford CD, Clark AM. Discovery and development of new drugs for systematic opportunistic infections, Atta-ur-Rahman (ed.), *Studies in Natural Product Chemistry*, Elsevier, Amsterdam, 2, 421-452, 1988.
14. Kirst HA, Michel KH, Mynderase JS, Chio EH, Yao RC, Nakasukasa WM, Boeck LD, Occlowitz JL, Paschal JW, Deeter JB, Thompson GD. Discovery, isolation and structure elucidation of a family of the structurally unique, fermentation derived tetracyclic macrolides, *Synthesis and Chemistry of Agrochemicals*, 111, 214-225, 1992.
15. Wedge DE, Galindo JCG, Macias FA. Fungicidal activity of natural and synthetic sesquiterpene lactone analogs, *Phytochemistry*, 53, 747-757, 2000.
16. Wedge DE, Kuhajek JM. A microbioassay for fungicide discovery, *SAAS Bulletin of Biochemistry and Biotechnology*, 11, 1-7, 1998.
17. Espinel-Ingroff A, Kerkering TM. Spectrophotometric method of inoculum preparation for the in vitro susceptibility testing of filamentous fungi, *Journal of Clinical Microbiology*, 29, 393-394, 1991.
18. Homans AL, Fuchs A. Direct bioautography on thin layer chromatography as a method for detecting fungitoxic substances, *Journal of Chromatography*, 51, 327-329, 1970.
19. Osborne AE, Chase BR, Lunness P, Scott PR, Daniels MJ. An oat species lacking avenacin is susceptible to infection by *Gaeumannomyces graminis* var. *tritici*, *Physiological and Molecular Plant Pathology*, 45, 457-467, 1994.
20. Akdemir ZS, Tatli II, Bedir E, Khan IA. Aceylated iridoid glycosides from *Verbascum lasianthum* Boiss., *Turk. J. Chem.*, 28, 101-109, 2004.
21. Miyase T, Horikoshi C, Yabe S, Miyasaka S, Melek FR, Kusano G. Saikosaponin homologues from *Verbascum* spp. The structures of mulleinsaponins I-VII, *Chem. Pharm. Bull.*, 45, 2029-2033, 1997.
22. Yesilada E. The biological effects and usage of saponins-I, *Pharmazia-JTPA* 26, 57, 153, 1986.
23. Hiller K. New results on the structure and biological activity of triterpene saponins, Hostettmann K, Lea PJ (eds.), *Biologically Active Natural Products*, Oxford University Press, New York, 1987.