

Bioavailability File: Amlodipine

Mustafa Sinan KAYNAK^{*o}, Agata BOGACZ^{*,***}, Michał STELMASIŃSKI^{*,***}, Selma ŞAHİN^{**}

Bioavailability File: Amlodipine

Amlodipine (AML), a third-generation dihydropyridin, is a long-acting L-calcium channel blocker used in the treatment of hypertension and angina pectoris. It exerts its effects by blocking the voltage-dependent L-calcium channels and binding to both dihydropyridin and nondihydropyridin binding sites. AML is well absorbed (96%) after oral administration and its bioavailability is between 64-90%. Its volume of distribution is about 16 to 21 L/kg and protein binding is 98% after oral administration. AML is extensively metabolized in the liver and its elimination from the plasma is biphasic with a terminal half-life of 30 to 50 h. It is excreted by renal route about 60%. According to Biopharmaceutics Classification System, AML is classified as class I drug by WHO. In this review physicochemical properties, pharmacology, analytical methods, pharmacokinetics and bioavailability of amlodipine are discussed.

Key Words: Amlodipine, Bioavailability, Pharmacokinetics, Biopharmaceutics Classification System (BCS)

Received: 10.03.2012

Revised: 26.11.2012

Accepted: 03.04.2013

Biyoyarlanım Dosyası: Amlodipin

Amlodipin (AML) yüksek tansiyon ve anjina pektoris tedavisinde kullanılan, üçüncü jenerasyon dihidropiridin türevi, uzun etkili L-kalsiyum kanal blokörü bir ilaçtır. Etkisini dihidropiridin ve nondihidropiridin bağlanma bölgelerine bağlanarak ve voltaja bağlı L-kalsiyum kanallarını bloke ederek göstermektedir. AML, oral uygulamadan sonra iyi absorplanmakta (%96) ve biyoyarlanımı %64 ile %90 arasında gerçekleşmektedir. Oral uygulamadan sonraki dağılım hacmi yaklaşık olarak 16-21 L/kg ve proteinlere bağlanması %98 olmaktadır. Amlodipin karaciğerde yüksek oranda metabolize olmakta ve plazmadan eliminasyonu iki fazlı olup eliminasyon yarılanma ömrü 30-50 saattir. Renal yolla %60 oranında itrah edilmektedir. AML Dünya Sağlık Örgütü tarafından biyofarmasötik sınıflandırma sistemine göre sınıf 1 ilaç olarak sınıflandırmaktadır. Bu derlemede amlodipinin fizyokimyasal özellikleri, farmakolojisi, analitik metodları, farmakokinetiği ve biyoyarlanımı tartışılmıştır.

Anahtar Kelimeler: Amlodipin, Biyoyarlanım, Farmakokinetik, Biyofarmasötik Sınıflandırma Sistemi (BSS)

* İnönü University, Faculty of Pharmacy, Department of Pharmaceutical Technology, 44280 MALATYA, TURKEY

** Hacettepe University, Faculty of Pharmacy, Department of Pharmaceutical Technology, 06100 Ankara, TURKEY

*** İnönü University, Faculty of Pharmacy, Undergraduate ERASMUS Exchange Student, 44280, MALATYA, TURKEY.

o Corresponding Author E-mail: mustafa.kaynak@inonu.edu.tr

INTRODUCTION

Amlodipine (AML), a third-generation dihydropyridin, is a long-acting L-calcium channel blocker used in the treatment of hypertension and angina pectoris (1-6). Like other calcium channel blockers AML causes relaxation of vascular smooth muscle and cardiac muscle (4-7). Pharmacokinetics of AML is very different from other drugs from its class (8-11). A pKa value of 8.7 means that AML is present in the ionized form at the physiologic pH. Therefore, it possesses a strong affinity for cell membranes. These characteristics are believed to be a reason for AML's unique pharmacokinetics (10, 11). AML has higher bioavailability, longer half-life ($t_{1/2}$), longer time to C_{max} , higher volume of distribution and slower gradual elimination than other calcium channel blockers (3, 4, 6, 8-12). Unique pharmacokinetic profile of AML is directly connected with clinical benefits (13). AML is the most frequently used antihypertensive drug among all dihydropyridines (14).

PHYSICOCHEMICAL PROPERTIES

Amlodipine besylate (AB) is a salt of AML (CAS 111470-99-6). Although it is used as a racemic mixture, only the S (-)-enantiomer is pharmacologically active, whereas R (+)-enantiomer is 1000-fold less active (2, 15). The chemical name is 3-ethyl-5-methyl(±)-2-[(2-aminoethoxy) methyl]-4-(2-chlorophenyl)-1,4-dihydro-6-methyl-3,5-pyridinedicarboxylate, monobenzenesulphonate. The empirical formula of AB is $C_{20}H_{25}ClN_2O_5 \cdot C_6H_6O_3S$ (1). The chemical structure of AB is shown in Figure 1.

AB is white to off-white, crystalline powder with a molecular weight of 567.06. It is slightly soluble in water and propanol, freely soluble in methanol, sparingly soluble in ethanol (1). Melting range of AB is 195-204°C. Theoretical and practical octanol/water partition coefficient ($\log K_{ow}$) are 3.00 and 2.66 respectively (16). AB is stable under ordinary conditions. AML has a pKa value of 8.7 (10, 11).

ANALYTICAL METHODS

Chromatography

Several HPLC methods have been described in the literature for determination of AML in human plasma (2, 3, 17-34), rat plasma (35), tablet and capsule (36)

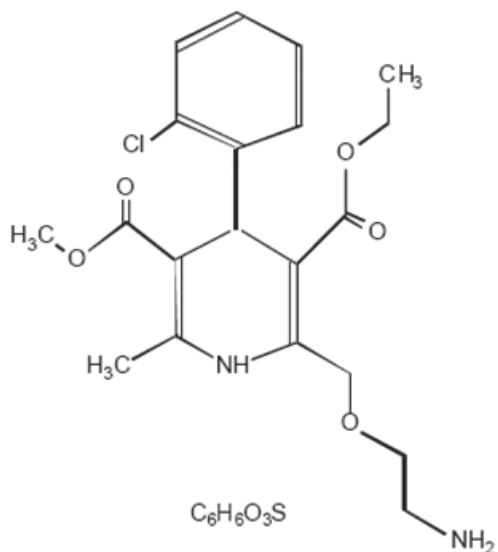


Figure 1. The chemical structure of AB (1).

samples (Table 1). Also various HPLC methods have been reported for determination of AML in combination with other drugs (37-46) (Table 2). All these methods differ with respect to the mobile phase, columns and detection methods used for the analysis of compounds. Other methods like thin layer chromatography (45), gas chromatography (47), high-performance thin-layer chromatography (48) were also developed to determine the amount of AML in human plasma or pharmaceutical formulations.

Spectroscopy

Several spectrophotometric methods have been described for determination of AML in pure form (49-52), pharmaceutical formulations (e.g. tablets, capsules) (36, 49, 51-53) and also for AML in combined pharmaceutical dosage forms (37, 54-56). Very few spectrofluorometric methods have appeared in literature for determination of AML in tablets (57, 58).

Voltammetry

Two voltammetric methods have been reported for determination of AML in human urine (59) and pharmaceutical formulations (59, 60).

PHARMACOLOGY

AML is a third-generation calcium channel antagonist, and inhibits the transmembrane influx of calcium ions into vascular smooth muscle and cardiac muscle. AML blocks voltage-dependent

Table 1. Chromatographic conditions of the reported methods used for the separation and determination of AML.

Samples	Column	Detection	Mobile Phase	Flow Rate (mL/min)	Extraction (+/-)	LOD/ LOQ (ng/mL)	Retation time (min)	Reference
Human plasma	HyPurity C18 (150x3.9 mm, 3µm)		ACN: 0.05M KH ₂ PO ₄ buffer: AA (62:38:01) (pH 3.5)	1.8		1.0/10		(17)
Human plasma	Bond Elut C2	Amperometric						(18)
Human plasma	C18 Column	Ultraviolet	10mM/L Ammonium Acetate: MeOH (30:70)	0.2	+	-/0.2		(19)
Human plasma	Supelcosil ABZ+Plus (25cmx4.6mm, 5µm)	Ultraviolet 360 nm	ACN: Water (70:30) with 10mM acetate buffer (pH 5)		+			(20)
Human plasma	Luna RP-C18 (15x2mm, 3µ)	Mass Spectrometry			+	>1.0/-		(21)
Human plasma	Monolithic WCX column				+	0.2/-		(22)
Human plasma	C18 Column (100x2.1mm, 3µm)	Mass Spectrometry	0.1%FA in ACN/0.1%FA in Water (42:58)	0.25	-	0.2/0.2	2.10	(3)
Human plasma	C18 Column	Mass Spectrometry			+		1.9	(23)
Human plasma	ACQUITY UPLC BEH C18 (50x2.1mm, 1.7µm)	Mass Spectrometry	Water: ACN (Both containing 0.3%FA)Gradient conditions	0.35		-/0.15		(24)
Human plasma	Nucleosil C8 (125x4.6mm)	Ultraviolet 239 nm	0.01 M NaH ₂ PO ₄ buffer :ACN (63:37) (pH 3.5)	1.5	+	0.2/-		(25)
Human plasma	Bondapak C18 (300x3.9mm, 10µm)	Fluorescence	MeOH: Water (80:20)	0.8	+		>20	(26)
Human plasma	X-Terra C18	Mass Spectrometry	0.02M Ammonium formate: ACN (20:80) (pH 4.5)	0.5			1.12	(27)
Human plasma	Chiral-AGP Column (150x10mm) Supelcosil LC8 (20x4.6mm, 5µm)		10mM Acetate buffer with 1% 1-propanolol (pH 4.5)	0.9	+	0.1/0.2	-	(2)
	Symmetry C8 (150x4.6mm, 4µm)	Ultraviolet 240 nm	10 mM Acetate buffer: ACN (55:45) (pH 4.5)					
Human plasma	Genesis C18 (150x4.6mm, 4µm)	Mass Spectrometry	60%CH ₃ CN: 40%Water + 10 mM FA	0.4	+	0.1/-	3.4	(28)
Human plasma		Mass Spectrometry						(29)
Human Plasma	C18 Column	Mass Spectrometry	MeOH:1% HAc (65:35)		+			(30)
Human Plasma	C18 Column	Fluorescence Exc: 470 nm Em: 537 nm	Sodium phosphate buffer with 1mL/L TEA: MeOH (pH 2.5)	2.8	+	-/0.25	>20	(31)
Human Plasma	Waters symmetry C18 (150x4.6, 5µm)	Mass Spectrometry	Water: ACN: FA (30:70:0.03)	1.0	+		1.5	(32)
Human Plasma	Hypersil BDS C18	Mass Spectrometry			+		3.2	(33)
Human plasma	C8 Column (150x4.6 mm, 5 µm)	Ultraviolet 238 nm	20 mM/L MeOH: KH ₂ PO ₄ (42:58) (pH 3.5)	1.0	+	0.1/-		(34)
Rat plasma		Electrochemical						(35)
Tablet Capsules	C18 Column		0.1% H ₃ PO ₄ : ACN (60:40) (pH 3.0)	1.0				(36)

Abbreviations: ACN: Acetonitrile, AA: Acetic Acid, MeOH : Methanol, FA: Formic Acid

Table 2. Chromatographic conditions of the reported methods used for the separation and determination of AML in combination with other drugs.

Samples	Column	Detection	Mobile Phase	Flow Rate (mL/min)	Extraction (+/-)	LOD/ LOQ (µg/mL)	Retention Time (min)	Reference
Binary mixtures (AML+VAL)	RP ACE C18 (150x4.6mm; 5µm)	Ultraviolet 254 nm	MeOH:ACN:NaH ₂ PO ₄ + 5mL/L TEA (42:18:40) (pH 3.0)	2.0				(37)
Commercial formulation (AML+RAM; AML+ ENA)		Ultraviolet 210 nm	MeOH:Water (50:50)					(38)
Rat liver perfusate containing AML and VAL	HICHROM Nucleosil 100-5 C18 (250x4.6mm)	Ultraviolet 240 nm	Phosphate buffer: ACN:MeOH (50:40:10) (0.01M, pH 3.6)	1.0	-	0.02/0.05	8.23	(39)
Tablet (AB+OLMED)	Kromasil C18 (250x4.6 mm)	Ultraviolet 238 nm	KH ₂ PO ₄ /K ₂ HPO ₄ ; ACN (50:50) (0.05 M)	1.0	-		3.69	(40)
Tablet (AB + BIS)	Luna C18-2 (50x4.6mm; 3µm)	Ultraviolet 230 nm	Ammonium buffer: ACN (65:35) (pH 5.0)	0.8	-	-	3.91	(41)
Tablet (AB+LOS+ HYDR)	Kromasil C18 (4.6 mm i.d.x250 mm)	Ultraviolet 232 nm	KH ₂ PO ₄ /K ₂ HPO ₄ :ACN (57:43) (0.025 M, pH=3.7)	1.0 (6.3min) 1.3 (6.3min)	-	-/-	5.12	(42)
Capsule (AB+BH)	Aquity UPLC, BEH C8 (100x2.1mm, 1.7µm)	Ultraviolet	Phosphate buffer: equal mix. ACN+MeOH (45:55) (pH 3.0)	0.3		-/0.01		(44)
Tablet (AB+VAL)	Zarbox ODS (4.6cmx250mm, 5µm)	Ultraviolet 254 nm	ACN: Phosphate buffer (50:50)	1.0	-	0.08/0.22	1.5	(93)
Human plasma Bulk powder Tablet (AB+VAL)	xTerra C18 (250x4.6mm, 5µm)	Ultraviolet 237 nm	MeOH: ACN: Water: 0.05%TEA (40:20:30:10) (pH 3.0 ±0.1)	1.2	-	-	-	(45)
Tablet dissolution samples (AML+VAL)	C18 ODS2 (200x4.6mm, 10µm)	Ultraviolet 240 nm	Phosphate Buffer:ACN:MeOH (44:46:10) (0.01 M, pH 3.6)	1.0	-	0.05/0.1	7.1	(46)

Abbreviations: AML: Amlodpine, AB: Amlodpine Besylate, VAL: Valsartan, RAM: Ramipril, ENA: Enalapril, OLMED: Olmesartan medoxomil, BIS: Bisoprolol fumarate, NH: Nebivolol Hydrochloride, LOS: Losartan Potassium, HYDR: Hydrochlorothiazide, BH: Benazepril Hydrochloride, ACN: Acetonitrile, MeOH : Methanol, TEA: Triethylamine, mix.: mixture.

L-calcium channels (1, 4, 6, 8, 10, 11, 61, 62) and it binds to both dihydropyridin and nondihydropyridin binding sites (1, 10). Because the contractile processes of muscles are dependent on amount of calcium ions in cells, the inhibition of calcium influx leads to vascular smooth muscle relaxation and negative inotropic and chronotropic effects in heart. AML acts selectively and it has greater effect on vascular smooth muscle cells than on cardiac

muscle cells (1, 4, 61, 62). Therefore, the negative inotropic effect is not significant *in vivo*, when AML is dosed in therapeutic amounts (1, 4, 10, 61, 62). Also the effects of AML on smooth muscle are more pronounced in arteries than in venous beds. AML causes reduction in peripheral resistance due to arterial dilatation and this is the main mechanism that leads to reduction in blood pressure and antianginal effects (1, 4, 61, 62) .

AML reduces supine and standing blood pressures. The heart rate and plasma catecholamine levels are not significantly changed when AML is chronically administered. With once daily administration the antihypertensive effect is maintained for at least 24 hours. In healthy, normotensive subjects, AML does not change blood pressure significantly (1) .

Wang et al. (63) investigated the effect of AB and dexamethasone combination in a gel formulation (0.5% and 0.3% respectively) on the ischemic skin flap. The results of the study showed that AB and dexamethasone in gel formulation might penetrate into skin tissue and could significantly increase the survival area of ischemic skin flap.

Side effects

Treatment with AB is usually well tolerated at doses up to 10 mg daily. The most common side effects are headache and edema. Other most common adverse reactions are: flushing, palpitation, fatigue, nausea, abdominal pain, and somnolence. The frequency and severity of adverse effects are connected with dose and several side effects (e.g. edema, flushing, palpitation, and somnolence) also with sex (more incidents in women than men). Other side effects such as cardiovascular problems, psychiatric problems, allergy, and musculoskeletal illnesses are very rare (the events occurred in less than 1% in placebo-controlled trials) (1, 64) .

Drug Interactions

In contrast to most of other calcium channel blockers, AML has few significant drug interactions. There is no clinically significant effect on the human plasma protein binding of digoxin, phenytoin, warfarin and indomethacin. Patients receiving drugs that induce or inhibit cytochrome P450 3A4 should be monitored for a potential change in AML response (1) .

FORMULATION TYPES

Although 2.5, 5 and 10 mg conventional tablets of AB (Norvasc®, Pfizer) are commercially available in the market, many researchers are trying to develop its rapidly dissolving tablets or dispersions with different preparation techniques and different excipients in order to increase patient compliance for

those who have trouble in swallowing tablets such as elderly or pediatric patients (1, 65-69) .

To investigate the stability of AML, Nahata et al. (70) prepared two suspension formulations containing 1 mg/mL AML using commercially available AML tablets (Norvasc-Pfizer). One of the formulation is in extemporaneously prepared 1% methylcellulose in syrup (1:1), and the other is in commercially available OraSweet®/OraPlus®. The results of the study showed that AML was stable in both suspension formulations stored in plastic prescription bottles for 91 days at 4°C or 56 days at 25°C. It was concluded that these formulations may be useful for elderly or pediatric patients who are unable to take tablets. Lyszkiewicz et al. (71) studied the bioavailability of Nahata et al.'s suspension formulations, and found that the bioavailability of the suspension formulations was not different from 5 mg tablet formulations. These findings support the use of the suspensions in children. Although, Nahata et al. claimed that the suspension formulations were stable for 56 days at room temperature, Lyszkiewicz et al. recommended freezing the suspensions during the using period or using it within a shorter period of time (70, 71) .

Various dosage forms of AB were investigated in the literature. The nanoemulsion drug delivery system of AB was designed by Chhabra et al. (72) to improve solubility and oral bioavailability of the drug and to localized delivery of drug at target size. Swamy et al. (73) were prepared intranasal hydroxypropyl guar (HPG) microspheres containing AB by using water in oil emulsification solvent evaporation technique in order to avoid first pass metabolism, to achieve controlled blood level profile and to improve therapeutic efficacy. Based on the results, it is suggested that, HPG is a suitable biodegradable polymer for nasal drug delivery of drugs with first pass metabolism such as AB. Asymmetric membrane capsules (AMCs) containing both AB and atenolol were prepared by Garg et al. (74). It was reported that the best AMCs formulation which consist of highest amount of osmotic agents and optimum amount of buffering agents followed zero order release kinetics for AB.

According to the FDA, AB has 184 drug products either alone or in combination with other drug (s). These drug products are registered as oral tablet or capsule dosage forms in combination of AB: Aliskiren Hemifumarate, AB:Atorvastatin Calcium, AB:Benazeprile Hydrochloride, AB:Olmesartan Medoxomil, AB:Telmisartan, AB:Valsartan, AB:Aliskiren Hemifumarate:Hydrochlorothiazide, AB:Olmesartan Medoxomil:Hydrochlorothiazide, AB:Valsartan:Hydrochlorothiazide (75) .

DOSAGE AND ADMINISTRATION

AB is available commercially in 2.5, 5 and 10 mg tablets. In adults the initial dose is 2.5 or 5 mg daily, the maximum dose is 10 mg daily. In pediatric patients, for 6-17 years of age, the effective antihypertensive dose is 2.5-5 mg once daily. The use of more than 5 mg once daily has not been studied in pediatric patients (1) .

PHARMACOKINETICS AND BIOAVAILABILITY

Absorption

AML is a dihydropyridin that has a slow absorption and prolonged effect. The extent of AML absorption is about 96% following oral administration (76). Following oral and i.v. doses of ¹⁴C-AML to rat and dog, 40-50% of the dose was excreted in the urine indicating that the oral dose was well absorbed (34). According to the “Martindale The Extra Pharmacopoeia” the bioavailability varies but is usually about 60 to 65% (77). When ¹⁴C-AML was administered to two human volunteers by means of single oral and intravenous doses, the drug was well absorbed by the oral route and the mean oral bioavailability for unchanged drug was 62.5% (76). Maximum plasma level is reached 6-12 hours after single oral administration, and absolute bioavailability of the AML tablet is estimated to be 64 to 90% (1, 9, 76, 78, 79) .

Faulkner et al. (79) investigated the pharmacokinetics of AML following single (10 mg, n=12, oral and IV) and repeated dose (15 mg/daily, n=28, 14 days, oral) administrations. The pharmacokinetic parameters determined after single and repeated dose administrations were given in Table 3. Comparative

Table 3. Pharmacokinetic parameters of AML obtained after single and repeated dose administrations (75).

Pharmacokinetic Parameters	Single dose (10 mg)	
	Oral	IV
AUC _{0-∞} (ng.h./mL)	238±53	371±69
C _{max} (ng/mL)	5.9±1.2	-
t _{max} (h)	7.6±1.8	-
k _{el} (h ⁻¹)	0.020±0.0036	0.021±0.0032
CL (mL/min per kg)	-	7.0±1.3
t _{1/2} (h)	35.7±6.1	33.8±5.3
V (L/kg)	-	21.4 + 4.4
Bioavailability (%)	64 Range 52-88	-
	Repeated oral dose	
	Day 1	Day 14
C _{max} (ng/mL)	6.9 ± 2.6	18.1 ± 7.1
t _{max} (h)	8.9 ± 3.7	8.7± 1.9
C _{min} (ng/mL)	3.3 ± 1.2	11.8 ± 5.3
C _{av} (ng/mL)	4.5 ± 1.6	14.5 ± 5.8
t _{1/2} (h)	-	44.7 8.6
k _{el} (1/h)	-	0.016 0.0034

pharmacokinetics after single iv and oral dose showed that bioavailability of oral AML is 64%. In repeated oral administration (once daily for 14 days, 15 mg), the steady state plasma drug concentration was reached by the seventh dose. Relatively long elimination half-life of AML (45 h) after repeated doses resulted in an approximately threefold accumulation.

Pharmacokinetic parameters of different AML salts (5, 80, 81) or generic AML tablets (78, 82) were examined in several studies. All studies showed no significant differences between pharmacokinetic and pharmacodynamic characteristics among different AML formulations. Carvalho et al. (28) investigated the bioequivalence of 5 mg AML (test formulation) or AB (reference formulation, Norvasc®) tablets in 24 healthy volunteers (Figure 2, Table 4). The study was conducted using an open-label, randomized

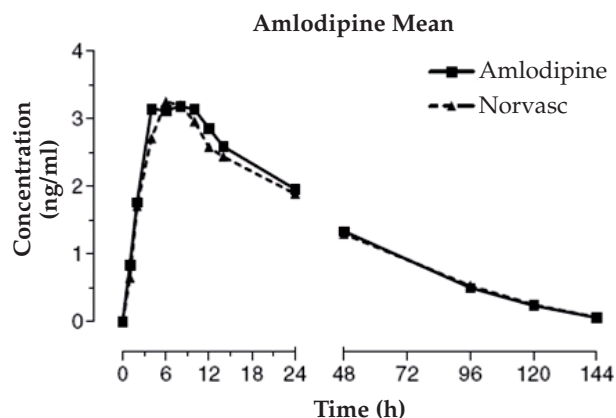


Figure 2. Mean plasma concentrations versus time curve for both AML formulations (28) .

two-period crossover design. It was found that AUC_{last} , AUC_{0-inf} and C_{max} ratios were within the 80–125% interval indicating that 5 mg AML tablet was bioequivalent to 5 mg Norvasc® tablet (28). Similarly, Liu et al. (3) was reported no significant difference between the pharmacokinetic parameters of test (dispersible AML tablet) and reference (Norvasc®) AML tablets obtained after oral administration of single dose (2x5 mg tablet) to healthy Chinese male volunteers.

Pharmacokinetics of AB in an AB/atorvastatin calcium (AC) combination tablet was investigated in a randomized, 2-way crossover design in 126 healthy volunteers. Subjects received a single dose of AB/AC tablet or coadministered matching doses of individual AB and AC tablets at the highest (10/80 mg; n=62) and the lowest (5/10 mg; n=64) dose strengths. The results of the study (Table 5 and Figure 3) demonstrated that there was no significant difference between the pharmacokinetic parameters obtained after oral administration of combination AB/AC tablets and coadministered individual AB and AC tablets (47) .

Chhabra et al. (72) were developed nanoemulsion (NE) of AB by spontaneous emulsification method with the aim to enhance the solubility and oral bioavailability of AB and to achieve localized drug delivery at target site. The drug release from NEs was significantly higher than the marketed tablet formulation ($p < 0.01$). The pharmacokinetics and

Table 4. Mean pharmacokinetic parameters obtained from 24 volunteers after oral administration of 5 mg AML tablet (28).

Pharmacokinetic parameter	AML Tablet	Norvasc® Tablet
AUC_{0-last} (ng.h/mL)		
Geometric Mean	151.7	147.4
S.D.	78.1	75.1
AUC_{0-inf} (ng.h/mL)		
Geometric Mean	166.9	166.3
S.D.	78.8	76.7
C_{max} (ng/mL)		
Geometric Mean	3.9	3.8
S.D.	2.5	2.1
k_e (1/h)		
Median	0.02	0.02
Range	0.01-0.03	0.01-0.04
$t_{1/2}$ (h)		
Median	33.9	37.0
Range	24.3-45.7	18.7-63.4
t_{max} (h)		
Median	6.0	6.0
Range	2.0-14.0	4.0-14.0

biodistribution studies of the optimized radiolabeled (99mTc-labeled) formulation (15% Labrafil M, 35% Tween 80: ethanol (2:1), and 50% by weight aqueous phase) in mice (p.o.) demonstrated a relative bioavailability of 475% against AB suspension. In almost all the tested organs, the uptake of AB from NE was significantly higher ($p < 0.05$) than AB suspension especially in heart with a drug targeting index of 44.1%, also confirming the efficacy of nanosized formulation at therapeutic site. A three times increase in the overall residence time of NE further signifies the advantage of NEs as drug carriers for enhancing bioavailability of AB (72) .

Distribution

AML has volume of distribution of 16 to 21 L/kg following oral administration (4, 83, 84). Tissue distribution is extensive in particular into the liver (84). AML is approximately 98% bound to plasma proteins in hypertensive patients (1, 83). The effect

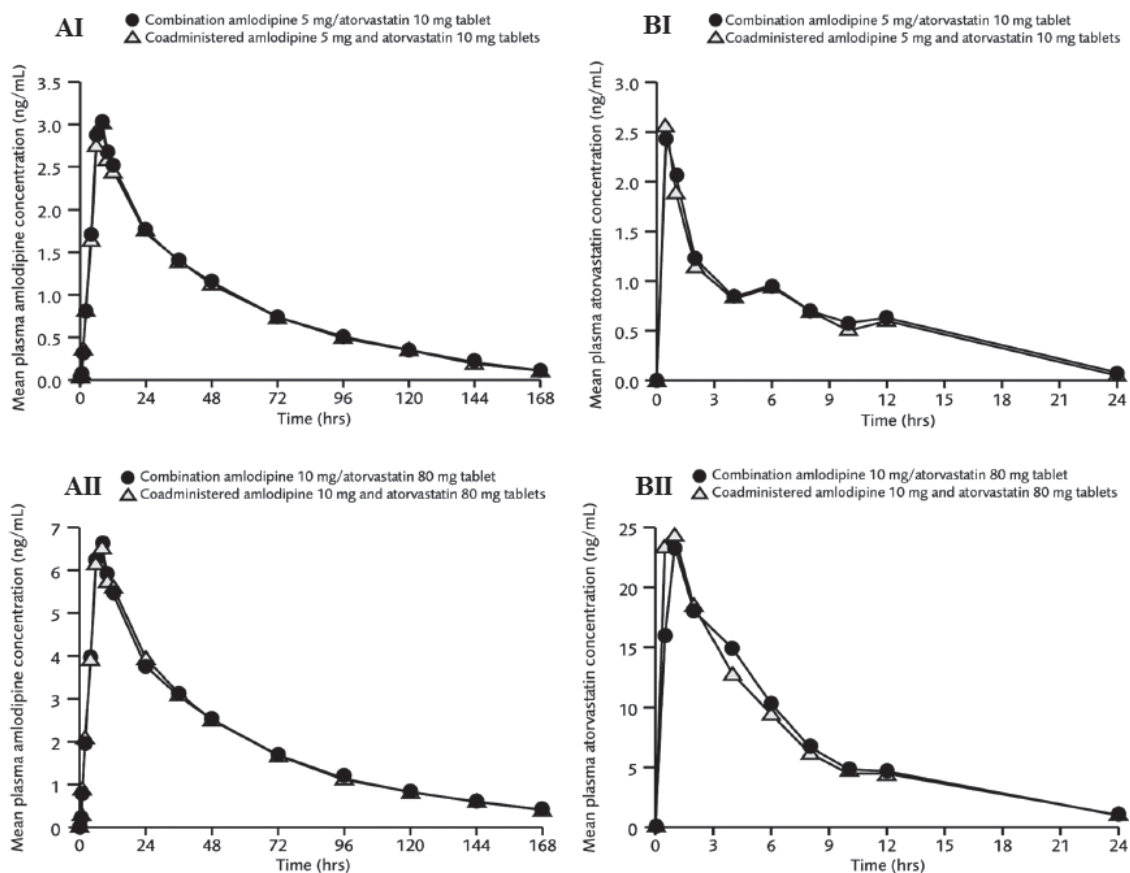


Figure 3. Mean plasma concentrations of AML (AI) and atorvastatin (BI) after the administration of 5/10-mg combination tablets versus the coadministration of AML 5-mg and atorvastatin 10-mg tablets. Mean plasma concentrations of AML (AII) and atorvastatin (BII) after the administration of 10/80-mg combination tablets versus the coadministration of AML 10-mg and atorvastatin 80-mg tablets (47).

Table 5. Mean (n = 63) Pharmacokinetic parameters obtained after oral administration of AML/Atorvastatine Calcium (AML/AC) combination tablets (5/10 mg or 10/80 mg) versus coadministration of individual AML (5 or 10 mg) and AC (10 or 80 mg) tablets (47).

	AML				AC			
	AUC _{0-inf} (ng.h/ mL)	C _{max} (ng/mL)	t _{max} (h)	t _{1/2} (h)	AUC _{0-inf} (ng.h/mL)	C _{max} (ng/mL)	t _{max} (h)	t _{1/2} (h)
5/10 mg combination tablet	151a	3.04a	7.80	44.9	16.2a	2.40a	0.79	7.30
AML 5 mg + AC 10 mg	147a	2.94a	7.67	45.1	15.6a	2.43a	0.81	7.60
Ratio of geometric mean, %	102.7	103.4	-	-	103.8	98.8	-	-
90% confidence interval	98.9-105.4	99.6-107.7	-	-	96.4-111.8	88.3-110.6	-	-
10/80 mg combination tablet	336a	6.63	7.61	45.8	163a	25.5a	0.89	9.10
AML 10 mg + AC 80 mg	336a	6.58a	8.07	46.9	156a	27.1a	1.54	9.34
Ratio of geometric mean, %	100.0	100.8	-	-	104.5	94.1	-	-
90% confidence interval	97.2-102.9	97.6-103.9	-	-	98.8-110.8	84.6-104.4	-	-

^a Geometric mean

of grapefruit juice on the pharmacokinetics of AML was investigated by Vincent et al. Water or grapefruit juice (240 mL) was administered to each subject prior to AML infusion. Amlodipine Maleate (10 mg) was administered via intravenous route and samples were collected for 216 hours from 20 male subjects. The calculated $V_{d_{ss}}$ values of water (control) and grapefruit juice (test) were 21.0 (± 3.8) and 22.7 (± 5.1) L/kg, respectively, with no significant difference between the groups (85).

Metabolism and Elimination

AML is extensively metabolized in the liver and its elimination from the plasma is biphasic with a terminal half-life of 30 to 50 h (1, 4, 6, 8-10, 83, 84). The rate of oxidative metabolism is relatively slow, therefore, AML does not exhibit extensive first-pass or presystemic metabolism after oral administration (83). AML is extensively (about 90%) converted to inactive metabolites via hepatic metabolism (Cytochrome P450 3A4 isozyme) (1).

Following oral and IV doses of ^{14}C -AML to rat and dog, urinary and faecal excretion in rat was essentially complete within 48 h but was prolonged upto 168 h in dog. The majority (about 95%) of the urinary metabolites were identified for both species; unchanged drug accounted for 10% and 20% of the urinary radioactivity in rat and dog respectively. In rat, the principal route of metabolism involved cleavage of the S-methoxycarbonyl group of both the parent dihydropyridine and its pyridine analogue. In contrast, metabolism in dog involved oxidative deamination of the 2-aminoethoxy-methyl side chain. Secondary metabolism in both rat and dog was similar to that of other calcium channel blockers of the dihydropyridine class, with oxidation to the pyridine form being followed by aliphatic hydroxylation in the 6-position or O-dealkylation in the 2-position and lactonization (34).

The disposition of AML, has been studied by Beresford et al. (76) in two human volunteers using single oral and IV doses of ^{14}C -AML. It was found that renal elimination was the major route of excretion with about 60% of the dosed radioactivity recovered in urine. Mean total recovered radioactivity in urine

and faeces amounted to 84% for both the oral and intravenous routes. Apart from a small amount of unchanged AML (10% of urine ^{14}C), only pyridine metabolites of AML were excreted in urine. Nine different metabolites of AML were identified (Figure 4). The major metabolite was 2- ([4- (2-chlorophenyl)-3-ethoxycarbonyl-S-methoxycarbonyl-6-methyl-2-pyridyl] methoxy) acetic acid (Figure 4, Met VII) and this represented 33% of urinary radioactivity. The majority (>95%) of these metabolites were excreted in the 0-72 h post-dose period. The data indicate that oxidation of AML to its pyridine analogue is the principal route of metabolism with subsequent metabolism by oxidative deamination, de-esterification and aliphatic hydroxylation. For the two volunteers, AML concentrations in plasma declined with a mean half-life of 33 h, while slower elimination of total drug-related material from plasma was observed, consistent with prolonged excretion (up to 12 days) of metabolites in urine and faeces. Only AML and pyridine metabolites were found in the circulation. As these pyridine derivatives have minimal calcium antagonist activity, the efficacy of AML in man can be attributed to the parent drug (76).

Food Effect

Absorption of AML is not affected by food (1, 15). Josefsson et al. investigated the effect of grapefruit juice on the pharmacokinetics of AML (5 mg single oral dose) in twelve healthy male volunteers. A single oral dose of AML (5 mg) was administered with a glass of grapefruit juice (250 mL) or water. When AML was coadministered with grapefruit juice, C_{max} (115%) and AUC (0-72 h; 116%) values were comparable with water, and no significant difference between t_{max} values (86). Similar observations were made by Vincent et al. (82). Single dose of oral and intravenous AML (10 mg) was administered to 20 healthy male volunteers. For 9 days beginning with the day of administration of AML, grapefruit juice (or water control) was given once daily, and blood samples, blood pressure and heart rate measures were obtained. Results of the study showed that oral AML has high systemic availability (grapefruit juice: 88%; water: 81%), and pharmacokinetic parameters (AUC, C_{max} , t_{max} , and k_{el}) were not markedly changed with grapefruit juice coadministration. Total plasma

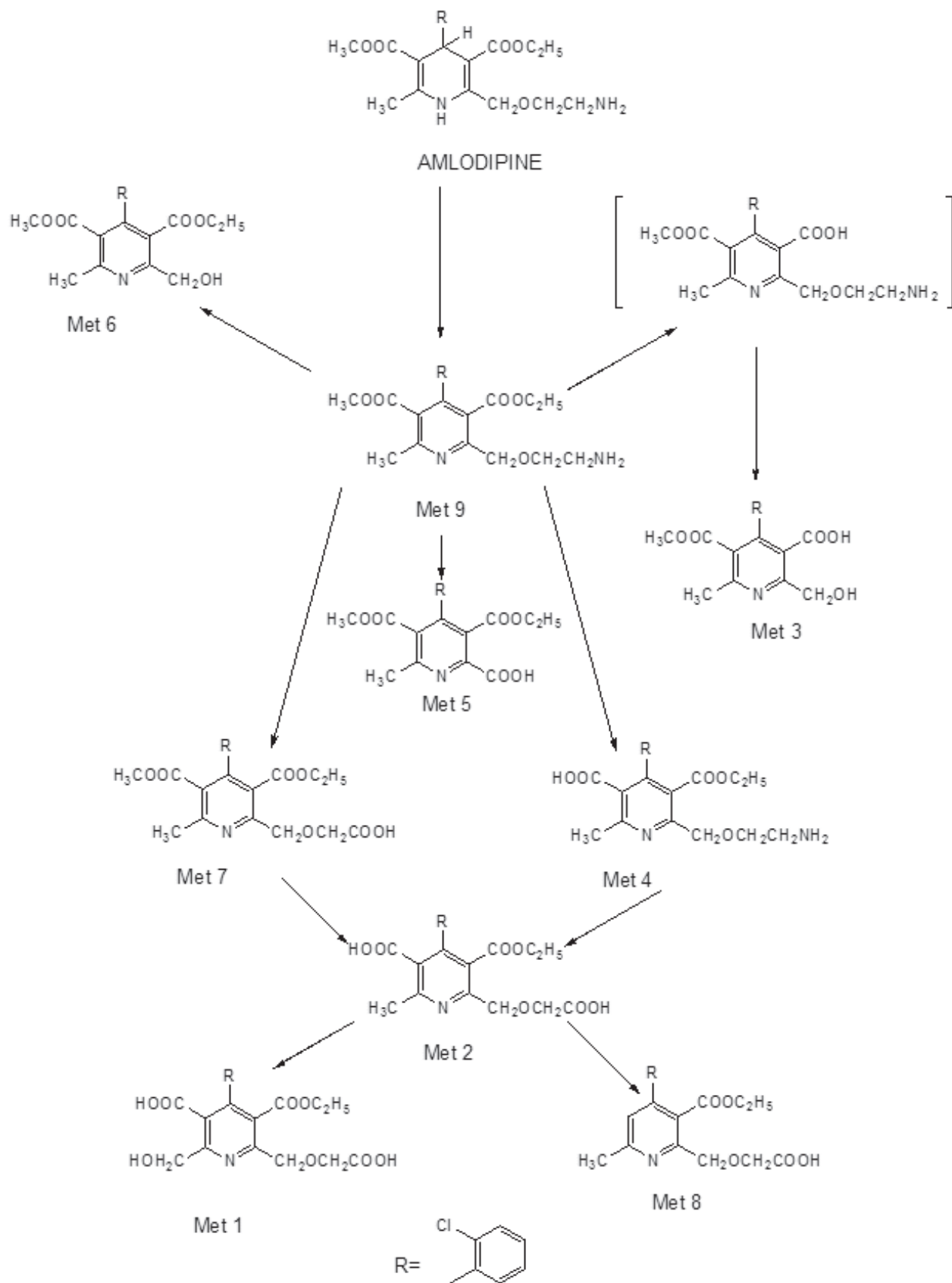


Figure 4. Urinary metabolites of AML in man (Met: Metabolite) (73).

clearance and volume of distribution, calculated after intravenous AML, were essentially unchanged by grapefruit juice (CL 6.65 mL/ min per kg, juice vs 6.93 mL/min per kg, water; $V_{d_{ss}}$ 22.7 L/ kg, juice vs 21.0 L/ kg, water). Also, grapefruit juice coadministration did not greatly alter the stereoselectivity in AML oral or intravenous kinetics. It was concluded that once daily grapefruit juice administration with usual oral doses of AML is unlikely to alter the profile of response in clinical practice (85) .

Renal impairment

The pharmacokinetics of AML is not significantly influenced by renal impairment and the dosage adjustment is not necessary (1, 87-89). When AML (as a single 5 mg capsule) was administered once daily for 14 days to 27 male subjects with renal functions ranging from normal to dialysis dependent (87), half-life and accumulation of AML were similar to previously reported values and did not vary with renal function. Similar observations were made when AML was administered (2.5-5.0 mg, once daily for 8 weeks) to 35 hypertensive patients with renal dysfunction (88) .

Hepatic impairment

Patients with hepatic insufficiency may require a lower initial dose of AML than healthy patients (1). When a single oral dose of AML (5 mg) was administered to 12 patients with hepatic impairment and 8 healthy convalescing subjects, some of the pharmacokinetic parameters were different in both groups. T_{max} was shorter and $t_{1/2}$ was longer in patients with hepatic insufficiency. Although AUCs were higher in hepatic patients, these differences were not significant. On the other hand, C_{max} values were similar in both groups (90) .

Age

The pharmacokinetic parameters of AML in children are not significantly different than those in adults and are not influenced by frequency of dosing (1). On the other hand, elderly patients have decreased clearance and longer $t_{1/2}$ suggesting increased drug accumulation during chronic dosing (1, 91) indicating that those patients may require a lower initial dose of AML (1) .

BIOPHARMACEUTICS CLASSIFICATION SYSTEM EVALUATION

The Biopharmaceutics Classification System (BCS) is a scientific framework for classifying drug substances based on their aqueous solubility and intestinal permeability. BCS categorizes drugs into four groups, Class 1 (high solubility and high permeability), Class 2 (low solubility and high permeability), Class 3 (high solubility and low permeability) and Class 4 (low solubility and low permeability). The BCS allows biowaiver for rapid dissolving immediate-release (IR) products of Class 1 drugs (92). On the other hand EMA extends biowaivers to drugs of Class 3 (high solubility and low permeability) (93). In regard to BCS classification of AML, there is conflicting information in "Proposal to Waive in Vivo Bioequivalence Requirements for the Who Model List of Essential Medicines Immediate Release, Solid Oral Dosage Forms". In Table A, AML is classified as BCS Class 1 compound. In Table C, its solubility is given as slightly soluble (5ml) and permeability as BAabs 60-65%, excretion of drug metabolites urine 90-95% (94). AML was classified as BCS Class 1 drug by Olusola et al and Shohin et al (95, 96) .

CONCLUSIONS

AML, which is a dihydropyridin derivative, has a slow absorption and prolonged effect. Following oral administration the extent of AML absorption is about 96%. AML is present in ionized form at physiologic pH because of its pKa value. Therefore, it possesses a strong affinity for cell membranes and has high permeability according to BCS guidance. These characteristics are believed to be a reason for AML's unique pharmacokinetics. AML has higher bioavailability, longer half-life ($t_{1/2}$), longer time to C_{max} , higher volume of distribution and slower gradual elimination than other calcium channel blockers. Because of its unique pharmacokinetics characteristics AML is the most frequently used antihypertensive drug among all dihydropyridines. AML pharmacokinetics is not significantly affected by co-administration with orange juice, age, renal impairment, hepatic impairment or age. According to some researchers and *WHO Model List of Essential Medicines* it was categorized as BCS Class 1 drug. Although it is used as a racemic mixture, only the S

(-)-enantiomer is pharmacologically active, whereas R (+)-enantiomer is 1000-fold less active. In some clinical studies, S-Amlodipine 2.5 mg is found to be equivalent in its efficacy and tolerability when compared to Amlodipine 5 mg in the treatment of mild to moderate hypertension.

REFERENCES

1. Norvasc (amlodipine besylate): Information of Health Care Professionals. Access Date: 29 September 2011. Available from: <http://labeling.pfizer.com/ShowLabeling.aspx?id=562>.
2. Luksa J, Josic D, Kremser M, Kopitar Z, Milutinovic S. Pharmacokinetic behaviour of R-(+)-and S-(-)-amlodipine after single enantiomer administration. *J Chromatogr B Biomed Sci Appl* 703 (1-2): 185-193, 1997.
3. Liu Y, Jia J, Liu G, Li S, Lu C, Yu C. Pharmacokinetics and bioequivalence evaluation of two formulations of 10-mg amlodipine besylate: an open-label, single-dose, randomized, two-way crossover study in healthy Chinese male volunteers. *Clin Ther* 31 (4): 777-783, 2009.
4. Scholz H. Pharmacological aspects of calcium channel blockers. *Cardiovasc Drugs Ther* 10 Suppl 3: 869-872, 1997.
5. Park JY, Kim KA, Park PW, Lee OJ, Kim JS, Lee GH, Ha MC, Park JH, O MJ, Ryu JH. Comparative pharmacokinetic and pharmacodynamic characteristics of amlodipine besylate and amlodipine nicotinate in healthy subjects. *Int J Clin Pharmacol Ther* 44 (12): 641-647, 2006.
6. Murdoch D, Heel RC. Amlodipine. A review of its pharmacodynamic and pharmacokinetic properties, and therapeutic use in cardiovascular disease. *Drugs* 41 (3): 478-505, 1991.
7. Derosa G, Maffioli P. Drug safety evaluation of amlodipine. *Expert Opin Drug Saf* 10 (5): 795-804, 2011.
8. Abernethy DR. Pharmacokinetics and pharmacodynamics of amlodipine. *Cardiology* 80 Suppl 1: 31-36, 1992.
9. Abernethy DR. The pharmacokinetic profile of amlodipine. *Am Heart J* 118 (5 Pt 2): 1100-1103, 1989.
10. Burges R, Moisey D. Unique pharmacologic properties of amlodipine. *Am J Cardiol* 73 (3): 2A-9A, 1994.
11. van Zwieten PA. Amlodipine: an overview of its pharmacodynamic and pharmacokinetic properties. *Clin Cardiol* 17 (9 Suppl 3): III3-6, 1994.
12. Chung M, Calcagni A, Glue P, Bramson C. Effect of food on the bioavailability of amlodipine besylate/atorvastatin calcium combination tablet. *J Clin Pharmacol* 46 (10): 1212-1216, 2006.
13. Meredith PA, Elliott HL. Amlodipine; clinical relevance of a unique pharmacokinetic profile. *J Cardiovasc Pharmacol* 22 Suppl A: S6-8, 1993.
14. Song YJ, Li J, Xie XF, Wang H, Li QX. Effects of amlodipine on TGF-beta-induced Smad2, 4 expressions in adriamycin toxicity of rat mesangial cells. *Arch Toxicol* 85 (6): 663-668, 2011.
15. Kim SA, Park S, Chung N, Lim DS, Yang JY, Oh BH, Tahk SJ, Ahn TH. Efficacy and safety profiles of a new S (-) -amlodipine nicotinate formulation versus racemic amlodipine besylate in adult Korean patients with mild to moderate hypertension: an 8-week, multicenter, randomized, double-blind, double-dummy, parallel-group, phase III, noninferiority clinical trial. *Clin Ther* 30 (5): 845-857, 2008.
16. Patel HJ, Patel JS, Desai BG, Patel KD. Permeability studies of anti hypertensive drug amlodipine besylate for transdermal delivery. *Asian J Pharm Clin Res* 3 (1): 31-34, 2010.
17. Alsarra IA. High-performance liquid chromatographic method for quantitative determination of amlodipine in human plasma and pharmaceutical dosage form and its application to pharmacokinetic studies. *J Chromatogr Sci* 47 (10): 863-867, 2009.
18. Josefsson M, Zackrisson AL, Norlander B. Sensitive high-performance liquid chromatographic analysis of amlodipine in human plasma with amperometric detection and a single-step solid-phase sample preparation. *J Chromatogr B Biomed Appl* 672 (2): 310-313, 1995.
19. Zou Q, Zhan Y, Ge Z, Wei P, Ouyang P. Liquid chromatography-mass spectrometry method for the determination of amlodipine in human plasma and its application in a bioequivalence study. *Arzneimittel-Forsch* 59 (8): 383-391, 2009.
20. Baranda AB, Etxebarria N, Jimenez RM, Alonso RM. Development of a liquid-liquid extraction procedure for five 1,4-dihydropyridines calcium

- channel antagonists from human plasma using experimental design. *Talanta* 67 (5): 933-941, 2005.
21. Baranda AB, Mueller CA, Alonso RM, Jimenez RM, Weinmann W. Quantitative determination of the calcium channel antagonists amlodipine, lercanidipine, nitrendipine, felodipine, and lacidipine in human plasma using liquid chromatography-tandem mass spectrometry. *Ther Drug Monit* 27 (1): 44-52, 2005.
 22. Wei X, Yang G, Qi L, Chen Y. Determination of nicardipine and amlodipine in human plasma using on-line solid-phase extraction with a monolithic weak cation-exchange column. *Talanta* 77 (3): 1197-1202, 2009.
 23. Massaroti P, Moraes LA, Marchioretto MA, Cassiano NM, Bernasconi G, Calafatti SA, Barros FA, Meurer EC, Pedrazzoli J. Development and validation of a selective and robust LC-MS/MS method for quantifying amlodipine in human plasma. *Anal Bioanal Chem* 382 (4): 1049-1054, 2005.
 24. Ma Y, Qin F, Sun X, Lu X, Li F. Determination and pharmacokinetic study of amlodipine in human plasma by ultra performance liquid chromatography-electrospray ionization mass spectrometry. *J Pharm Biomed Anal* 43 (4): 1540-1545, 2007.
 25. Zarghi A, Foroutan SM, Shafaati A, Khoddam A. Validated HPLC method for determination of amlodipine in human plasma and its application to pharmacokinetic studies. *Farmaco* 60 (9): 789-792, 2005.
 26. Tatar S, Atmaca S. Determination of amlodipine in human plasma by high-performance liquid chromatography with fluorescence detection. *J Chromatogr B* 758 (2): 305-310, 2001.
 27. Ramani AV, Sengupta P, Mullangi R. Development and validation of a highly sensitive and robust LC-ESI-MS/MS method for simultaneous quantitation of simvastatin acid, amlodipine and valsartan in human plasma: application to a clinical pharmacokinetic study. *Biomed Chromatogr* 23 (6): 615-622, 2009.
 28. Carvalho M, Oliveira CH, Mendes GD, Sucupira M, Moraes ME, De Nucci G. Amlodipine bioequivalence study: quantification by liquid chromatography coupled to tandem mass spectrometry. *Biopharm Drug Dispos* 22 (9): 383-390, 2001.
 29. Pico JC, Dominguez G, Negri AL, Caubet JC, Terragno NA. Comparative pharmacokinetics of a single oral dose of two formulations of amlodipine. A randomized, single-blind, two-period, two-sequence, crossover study. *Arzneimittel-Forsch* 58 (7): 323-327, 2008.
 30. Feng Y, Zhang L, Shen Z, Pan F, Zhang Z. Analysis of amlodipine in human plasma by liquid chromatography-mass spectrometry. *J Chromatogr Sci* 40 (1): 49-53, 2002.
 31. Bahrami G, Mirzaeei S. Simple and rapid HPLC method for determination of amlodipine in human serum with fluorescence detection and its use in pharmacokinetic studies. *J Pharm Biomed Anal* 36 (1): 163-168, 2004.
 32. Nirogi RV, Kandikere VN, Mudigonda K, Shukla M, Maurya S. Sensitive and rapid liquid chromatography/tandem mass spectrometry assay for the quantification of amlodipine in human plasma. *Biomed Chromatogr* 20 (9): 833-842, 2006.
 33. Bhatt J, Singh S, Subbaiah G, Shah B, Kambli S, Ameta S. A rapid and sensitive liquid chromatography-tandem mass spectrometry (LC-MS/MS) method for the estimation of amlodipine in human plasma. *Biomed Chromatogr* 21 (2): 169-175, 2007.
 34. Beresford AP, Macrae PV, Stopher DA. Metabolism of amlodipine in the rat and the dog: a species difference. *Xenobiotica* 18 (2): 169-182, 1988.
 35. Shimooka K, Sawada Y, Tatematsu H. Analysis of amlodipine in serum by a sensitive high-performance liquid chromatographic method with amperometric detection. *J Pharm Biomed Anal* 7 (11): 1267-1272, 1989.
 36. Malesuik MD, Cardoso SG, Bajerski L, Lanzanova FA. Determination of amlodipine in pharmaceutical dosage forms by liquid chromatography and ultraviolet spectrophotometry. *J AOAC Int* 89 (2): 359-364, 2006.
 37. Kul D, Dogan-Topal B, Kutucu T, Uslu B, Ozkan SA. High-Performance Liquid Chromatographic and First Derivative of the Ratio Spectrophotometric Determination of Amlodipine and Valsartan in

- Their Binary Mixtures. *J AOAC Int* 93 (3): 882-890, 2010.
38. Bhushan R, Gupta D, Singh SK. Liquid chromatographic separation and UV determination of certain antihypertensive agents. *Biomed Chromatogr* 20 (2): 217-224, 2006.
39. Çelebier M, Kaynak MS, Altınöz S, Sahin S. Validated HPLC method development: The simultaneous analysis of amlodipine and valsartan in samples for liver perfusion studies. *Hacettepe Univ J Fac Pharm* 28 (1): 15-30, 2008.
40. Wankhede SB, Wadkar SB, Raka KC, Chitlange SS. Simultaneous estimation of amlodipine besylate and olmesartan medoxomil in pharmaceutical dosage form. *Indian J Pharm Sci* 71 (5): 563-567, 2009.
41. Vora DN, Kadav AA. Development and Validation of a Simultaneous HPLC Method for Estimation of Bisoprolol Fumarate and Amlodipine Besylate from Tablets. *Indian J Pharm Sci* 70 (4): 542-546, 2008.
42. Wankhede SB, Raka KC, Wadkar SB, Chitlange SS. Spectrophotometric and HPLC methods for simultaneous estimation of amlodipine besylate, losartan potassium and hydrochlorothiazide in tablets. *Indian J Pharm Sci* 72 (1): 136-140, 2010.
43. Al-Shaalan NH, Alnowaiser MA. Simultaneous determination of Amlodipine besylate and Valsartan in pharmaceutical formulation using high performance liquid chromatography. *J Chem Pharm Res* 2 (6): 129-134, 2010.
44. Kasawar GB, Farooqui MN. Simultaneous Determination of Amlodipine Besylate and Benazepril Hydrochloride in Pharmaceutical Dosage Form by LC. *Anal Sci* 25 (12): 1495-1498, 2009.
45. Ramadan NK, Mohamed HM, Moustafa AA. Rapid and Highly Sensitive HPLC and TLC Methods for Quantitation of Amlodipine Besylate and Valsartan in Bulk Powder and in Pharmaceutical Dosage Forms and in Human Plasma. *Anal Lett* 43 (4): 570-581, 2010.
46. Çelebier M, Kaynak MS, Altınöz S, Sahin S. HPLC method development for the simultaneous analysis of amlodipine and valsartan in combined dosage forms and in vitro dissolution studies. *Braz J Pharm Sci* 46 (4): 761-768, 2010.
47. Chung MG, Calcagni A, Glue P, Bramson C. Bioavailability of amlodipine besylate/atorvastatin calcium combination tablet. *J Clin Pharmacol* 46 (9): 1030-1037, 2006.
48. Pandya KK, Satia M, Gandhi TP, Modi IA, Modi RI, Chakravarthy BK. Detection and Determination of Total Amlodipine by High-Performance Thin-Layer Chromatography—a Useful Technique for Pharmacokinetic Studies. *J Chromatogr B Biomed Appl* 667 (2): 315-320, 1995.
49. Rahman N, Hoda MN. Validated spectrophotometric methods for the determination of amlodipine besylate in drug formulations using 2,3-dichloro 5,6-dicyano 1,4-benzoquinone and ascorbic acid. *J Pharm Biomed Anal* 31 (2): 381-392, 2003.
50. Dangi M, Chaudhari D, Sinkar M, Racha V, Damle MC. Stability Indicating HPTLC Method for Estimation of Nebivolol Hydrochloride and Amlodipine Besylate in Combination. *Eurasian J Anal Chem* 5 (2): 161-169, 2010.
51. Basavaiah K, Chandrashekar U, Prameela HC. Sensitive spectrophotometric determination of amlodipine and felodipine using iron (III) and ferricyanide. *Farmaco* 58 (2): 141-148, 2003.
52. Rahman N, Azmi SNH. Spectrophotometric method for the determination of amlodipine besylate with ninhydrin in drug formulations. *Farmaco* 56 (10): 731-735, 2001.
53. Rahman N, Singh M, Hoda MN. Application of oxidants to the spectrophotometric determination of amlodipine besylate in pharmaceutical formulations. *Farmaco* 59 (11): 913-919, 2004.
54. Nagavalli D, Vaidhyalingam V, Santha A, Sankar ASK, Divya O. Simultaneous spectrophotometric determination of losartan potassium, amlodipine besylate and hydrochlorothiazide in pharmaceuticals by chemometric methods. *Acta Pharmaceutica* 60 (2): 141-152, 2010.
55. Patil PR, Rakesh SU, Dhabale PN, Burade KB. Simultaneous UV spectrophotometric method for estimation of Losartan Potassium and Amlodipine Besylate in tablet dosage form. *Asian J Research Chem* 2 (1): 183-187, 2009.
56. Ramesh D, Ramakrishna S. New spectrophotometric methods for simultaneous determination of Amlodipine Besylate and Atorvastatin

- Calcium in tablet dosage forms. *Int J Pharm Pharm Sci* 2 (4): 215-219, 2010.
57. Abdel-Wadood HM, Mohamed NA, Mahmoud AM. Validated spectrofluorometric methods for determination of amlodipine besylate in tablets. *Spectrochim Acta A Mol Biomol Spectrosc* 70 (3): 564-570, 2008.
58. Shaalan RA, Belal TS. Simultaneous spectrofluorimetric determination of amlodipine besylate and valsartan in their combined tablets. *Drug Test Anal* 2 (9-10): 489-493, 2010.
59. Goyal RN, Bishnoi S. Voltammetric determination of amlodipine besylate in human urine and pharmaceuticals. *Bioelectrochemistry* 79 (2): 234-240, 2010.
60. Ahtiokka G, Dogrukol-Ak D, Tuncel M, Aboul-Enin HY. Determination of Amlodipine in pharmaceutical formulations by differential-pulse voltammetry with a glassy carbon electrode. *Arch Pharm* 335 (2-3): 104-108, 2002.
61. Webster J, Robb OJ, Jeffers TA, Scott AK, Petrie JC, Towler HM. Once daily amlodipine in the treatment of mild to moderate hypertension. *Br J Clin Pharmacol* 24 (6): 713-719, 1987.
62. Burges RA, Dodd MG, Gardiner DG. Pharmacologic profile of amlodipine. *Am J Cardiol* 64 (17): 10I-18I; discussion 18I-20I, 1989.
63. Wang X, Zhang X, Qin Y, Zhong L, Liu K, Zhang J. Percutaneous penetration ability of dexamethasone-amlodipine besylate compound gel and its effect on survival of ischemic random skin flap. *Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi* 24 (5): 566-570, 2010.
64. Hong SJ, Ahn TH, Baek SH, Cho WH, Jeon HK, Kwan J, Yoon MH, Lee KJ, Lim DS. Comparison of efficacy and tolerability of amlodipine orotate versus amlodipine besylate in adult patients with mild to moderate hypertension: a multicenter, randomized, double-blind, placebo-controlled, parallel-group, 8-week follow-up, noninferiority trial. *Clin Ther* 28 (4): 537-551, 2006.
65. Gustafsson D, Nystrom J, Carlsson S, Bredberg U, Eriksson U, Gyzander E, Elg M, Antonsson T, Hoffmann K, Ungell A, Sorensen H, Nagard S, Abrahamsson A, Bylund R. The direct thrombin inhibitor melagatran and its oral prodrug H 376/95: intestinal absorption properties, biochemical and pharmacodynamic effects. *Thromb Res* 101 (3): 171-181, 2001.
66. Bhardwaj V, Shukla V, Goyal N, Salim MD, Sharma PK. Formulation and evaluation of fast disintegrating sublingual tablets of amlodipine besylate using different superdisintegrants. *Int J Pharmacy Pharm Sci* 2 (3): 89-92, 2010.
67. Pare A, Yadav SK, Patil UK. Formulation and evaluation of effervescent floating tablet of amlodipine besylate. *Res J Pharm and Tech* 1 (4): 526-530, 2008.
68. Narmada GY, Mohini K, Prakash RB, Gowrinath DXP, Kumar KS. Formulation, evaluation and optimization of fast dissolving tablets containing amlodipine besylate by sublimation method. *ARS Pharm* 50 (3): 129-144, 2009
69. Bhardwaj V, Mayank B, Sharma PK. Formulation and evaluation of fast dissolving tablets of amlodipine besylate using different super disintegrants and camphor as sublimating agent. *Am-Euras J Sci Res* 5 (4): 264-269, 2010.
70. Nahata MC, Morosco RS, Hipple TF. Stability of amlodipine besylate in two liquid dosage forms. *J Am Pharm Assoc (Wash)* 39 (3): 375-377, 1999.
71. Lyszkiewicz DA, Levichek Z, Kozer E, Yagev Y, Moretti M, Hard M, Koren G. Bioavailability of a pediatric amlodipine suspension. *Pediatr Nephrol* 18 (7): 675-678, 2003.
72. Chhabra G, Chuttani K, Mishra AK, Pathak K. Design and development of nanoemulsion drug delivery system of amlodipine besylate for improvement of oral bioavailability. *Drug Dev Ind Pharm* 37 (8): 907-916, 2011.
73. Swamy NG, Abbas Z. Preparation and in vitro characterization of mucoadhesive hydroxypropyl guar microspheres containing amlodipine besylate for nasal administration. *Indian J Pharm Sci* 73 (6): 608-614, 2011.
74. Garg S, Pathak K, Philip A, Puri D. Osmotically regulated two-compartment asymmetric membrane capsules for simultaneous controlled release of anti-hypertensive drugs. *Sci Pharm* 80 (1): 229-250, 2012.
75. Approved Drug Products with Therapeutic Equivalence Evaluations. 10.02. Available from: <http://www.accessdata.fda.gov/scripts/cder/ob/docs/tempai.cfm>.

76. Beresford AP, McGibney D, Humphrey MJ, Macrae PV, Stopher DA. Metabolism and kinetics of amlodipine in man. *Xenobiotica* 18 (2): 245-254, 1988.
77. Martindale "The Extrapharmacopoeia", ed. S.C. Sweetman. Vol. 35th Ed. 2007, London, U.K.: Pharmaceutical Press.
78. Sailer R, Arnold P, Erenmemisoglu A, Martin W, Tamur U, Kanzik I, Hincal AA. Pharmacokinetics and bioequivalence study of a generic amlodipine tablet formulation in healthy male volunteers. *Arzneimittel-Forsch* 57 (7): 462-466, 2007.
79. Faulkner JK, McGibney D, Chasseaud LF, Perry JL, Taylor IW. The pharmacokinetics of amlodipine in healthy volunteers after single intravenous and oral doses and after 14 repeated oral doses given once daily. *Br J Clin Pharmacol* 22 (1): 21-25, 1986.
80. Kim BH, Kim JR, Kim MG, Kim KP, Lee BY, Jang IJ, Shin SG, Yu KS. Pharmacodynamic (hemodynamic) and pharmacokinetic comparisons of S-amlodipine gentisate and racemate amlodipine besylate in healthy Korean male volunteers: two double-blind, randomized, two-period, two-treatment, two-sequence, double-dummy, single-dose crossover studies. *Clin Ther* 32 (1): 193-205, 2010.
81. Mignini F, Tomassoni D, Traini E, Amenta F. Single-dose, randomized, crossover bioequivalence study of amlodipine maleate versus amlodipine besylate in healthy volunteers. *Clin Exp Hypertens* 29 (8): 539-552, 2007.
82. Rojanasthien N, Teekachunhatean S, Jakob K, Gaupp M, Arnold P, Chaichana N, Martin W. Bioequivalence study of generic amlodipine in healthy Thai male volunteers. *Int J Clin Pharmacol Ther* 42 (6): 330-335, 2004.
83. Meredith PA, Elliott HL. Clinical pharmacokinetics of amlodipine. *Clin Pharmacokinet* 22 (1): 22-31, 1992.
84. Walker DK, Humphrey MJ, Smith DA. Importance of metabolic stability and hepatic distribution to the pharmacokinetic profile of amlodipine. *Xenobiotica* 24 (3): 243-250, 1994.
85. Vincent J, Harris SI, Foulds G, Dogolo LC, Willavize S, Friedman HL. Lack of effect of grapefruit juice on the pharmacokinetics and pharmacodynamics of amlodipine. *Br J Clin Pharmacol* 50 (5): 455-463, 2000.
86. Josefsson M, Zackrisson AL, Ahlner J. Effect of grapefruit juice on the pharmacokinetics of amlodipine in healthy volunteers. *Eur J Clin Pharmacol* 51 (2): 189-193, 1996.
87. Doyle GD, Donohue J, Carmody M, Laher M, Greb H, Volz M. Pharmacokinetics of amlodipine in renal impairment. *Eur J Clin Pharmacol* 36 (2): 205-208, 1989.
88. Saruta T, Ishii M, Abe K, Iimura I. Efficacy and safety of amlodipine in hypertensive patients with renal dysfunction. *Clin Cardiol* 17 (6): 317-324, 1994.
89. Osterloh I. The safety of amlodipine. *Am Heart J* 118 (5 Pt 2): 1114-1120, 1989.
90. Darnis F, Poupon R. Pharmacokinetics and safety of single oral doses of amlodipine in patients with and without hepatic impairment: an open study. *Int J Clin Pharmacol Res* 13 (1): 29-33, 1993.
91. Abernethy DR, Gutkowska J, Lambert MD. Amlodipine in elderly hypertensive patients: pharmacokinetics and pharmacodynamics. *J Cardiovasc Pharmacol* 12 Suppl 7: S67-71, 1988.
92. FDA FDA "Waiver of In-Vivo Bioavailability and Bioequivalence Studies for Immediate-Release Solid Oral Dosage Forms based on a Biopharmaceutics Classification System", August 2000.
93. EMEA Guideline on the Investigation of Bioequivalence, 1 August 2010.
94. World Health Organization (WHO), Annex 8 Proposal to waive in vivo bioequivalence requirements for WHO Model List of Essential Medicines immediate-release, solid oral dosage forms. *WHO Technical Report Series* No: 937, 2006.
95. Olusola AM, Olubukola OO, Emeka OH, Lilian AE. Equivalence of Two Generic Brands of Amlodipine Besylate Under Biowaiver Conditions. *Int J Pharm Pharm Sci* 4 (2): 265-268, 2012.
96. Shohin IE, Ramenskaya GV, Vasilenko GF, Malashenko EA. In Vitro Dissolution Kinetics of Amlodipine Tablets Marketed in Russia Under Biowaiver Conditions. *Dissolut Technol* 17 (3): 20-22, 2010.