

A Study on the Optimization of CYP2E1 Enzyme Activity in C57Bl/6 Mouse Brain and Liver

Rahman BAŞARAN, Elçin Deniz ÖZDAMAR, Benay CAN EKE^o

A Study on the Optimization of CYP2E1 Enzyme Activity in C57Bl/6 Mouse Brain and Liver

C57Bl/6 Fare Beyin ve Karaciğer Mikrozomlarında CYP2E1 Enzim Aktivitesi Optimizasyon Çalışması

SUMMARY

In recent years, some studies have shown that CYP2E1 enzyme is associated with many diseases. Determining the activity of this enzyme will help to obtain new clues about several mechanisms in unknown progression of diseases. The purpose of this study is to assess the optimal conditions for CYP2E1 activity. In this context, the optimum conditions (amount of protein, substrate concentration and incubation time) were investigated in C57Bl/6 mouse brain and liver microsomes. In this study, it was found out that, different reaction conditions are required for this enzyme activities in microsomal brain and liver CYP2E1 of C57Bl/6 mice. According to our research results, these optimization studies should be examined for CYP2E1 activity in different tissues of mouse for future investigation.

Key Words: Brain, CYP2E1, CYP450, liver, mouse.

Received: 04.12.2014

Revised: 21.12.2014

Accepted: 22.01.2015

ÖZET

Son yıllarda yapılan araştırmalar, CYP2E1 enzimi ile birçok hastalığın ilişkisi olabileceğine dair veriler ortaya koymuştur. Bu enzim aktivitesinin belirlenmesi, hastalık süreçlerinde bilinmeyen birçok mekanizmaya ışık tutabilir. Bu çalışmanın amacı, CYP2E1 enzim aktivitesi için gerekli optimum koşulları (protein miktarı, substrat konsantrasyonu ve inkübasyon zamanı) araştırmak ve değerlendirmektir. Bu çalışmada, C57Bl/6 fare beyin ve karaciğer CYP2E1 enzim aktivitesi için farklı reaksiyon koşullarına ihtiyaç duyulduğu gözlenmiştir. Optimizasyon sonuçlarımız, daha kapsamlı araştırmalar için CYP2E1 enzim aktivitesinin farklı fare dokularında da çalışmasının gerekliliğini ortaya koymaktadır.

Anahtar kelimeler: Beyin, CYP2E1, CYP450, fare, karaciğer

INTRODUCTION

The most important enzyme system of phase I metabolism is cytochrome P450, a superfamily of heme-containing mono-oxygenases and responsible for the metabolism of drugs, xenobiotics, environmental compounds, carcinogens and several endogenous compounds (1,2). These enzymes are abundant in the liver, it also exists in some extrahepatic tissues such as kidney, lung, intestine and brain (4). Total CYP levels in the brain are low and approximately 0,5-2% of those in the liver (2). CYPs are distributed homogeneously in the liver whereas each of CYP450 isoenzymes is located according to their specific localization in the brain (3). Therefore, it is difficult to determine the enzymatic activity of CYPs in the brain.

CYP2E1 is involved in the detoxification-activation reactions of various endogenous and exogenous com-

pounds. Besides a role in the ethanol metabolism, CYP2E1 catalyses the metabolism and activation of drugs (e.g. halothane, enflurane, sevoflurane, acetaminophen), endogenous substrates (e.g. fatty acids, ketones), several procarcinogens (e.g. nitrosamines, benzene) and halogenated hydrocarbons (e.g. carbon tetrachloride, 1,1,1-trichloroethane, methlychloride, vinylchloride) (4-7).

It was the first identified in the microsomal ethanol-inducing system (MEOS) (8). This P450 isoform has been observed in many organism such as rabbit, rat, mice and human. CYP2E1 is one of the preserved forms in CYP450 enzymes family and its catalytic activities are quite similar among the species. It is difficult to find a common feature of substances that are metabolized by CYP2E1 but in general, small, hydrophobic and under 70 kDa molecular weight compounds (more

than 70 different chemicals) are suitable to be catalyzed by CYP2E1 (9,10).

Among CYP450 enzymes, CYP2E1 has a remarkable effect on toxicity. It can also generate reactive oxygen species during its catalytic cycle which contributes to oxidative stress and causing lipid peroxidation, protein inactivation and DNA damage (11). The substrates of CYP2E1 have become more toxic and carcinogenic substances via its catalytic activity (16). Thus, it is thought to have a role in progression of diseases. On the other hand, CYP2E1 levels in liver increases under some pathophysiological conditions such as alcohol consumption (12), diabetes (13), obesity (14) and starvation (15) in animals and humans.

As other CYPs, CYP2E1 levels are low in the brain when compared to the liver. But, most of the studies have indicated that activities of this enzyme can vary in the presence of inducer or inhibitor. Some regions and cells CYP2E1 levels in the brain are expressed as high as those in the liver. In experimental conditions, recent studies have shown inducer-dependent induction of brain CYP2E1 catalytic activity to be %125 of the liver both in rats and humans (17-22).

Its catalytic activity can be measured with N-nitrosodimethylamine (NDMA), p-Nitrophenol (pNP) and Chlorzoxazone (CLZ) *in vitro* (23). Based on previous studies, pNP was selected substrate of CYP2E1. This chemical has been indicated the activity of CYP2E1 in microsomal samples containing. pNP is a phenolic substance and a solution of the phenolate salt is yellow. CYP2E1 hydroxylates pNP at the 2 position to generate 4-Nitrocatechol (4NC) (24).

Activities of brain and hepatic CYPs are of key relevance for the susceptibility to system diseases and treatment outcomes. CYP2E1 is known to be expressed in mouse liver, as well as in various mouse brain regions. In this study, the catalytic activity of CYP2E1 can be estimated in C57Bl/6 brain and liver tissues, using p-nitrophenol metabolism. Determination of the CYP2E1 catalytic activity in brain and liver may be important from several toxicological points of view.

MATERIALS AND METHODS

Chemicals

Folin-ciocalteu reagent, Glucose-6-phosphate, Glucose-6-phosphate dehydrogenase, Nicotinamide adenine dinucleotide phosphate, sodium salt (NADP⁺), perchloride acid, p-nitrophenol, sodium hydroxide were purchased from Sigma-Aldrich (USA). All reagents were of analytical grade.

Animals

All procedures used in this study were approved by the Ankara University Animal Care and Use Committee.

C57Bl/6 (20-25 g, 12 weeks, male) mice were used throughout the experiments. Mice were provided a week ago, because travel changes the brain chemicals (25). The animals were housed 4 per cage in an animal house under standard conditions (12-h light-dark cycle, room temperature; 22-25 °C, optimum humidity) and fed standard pellet and water *ad libitum*. The animals were sacrificed via cervical dislocation. Brain and liver tissues were carefully dissected and immediately stored at -80 °C.

Preparation of Microsomes

The mouse brain and liver tissues were homogenized (%1,15 KCl (w/v)) in ice-cold condition, and centrifuged for 25 min. at 10.000 g. Centrifugation of the 10.000 g supernatants at 108.000 g. The pellets of ultracentrifugation were homogenized with %20 glycerol and were stored at -80 °C until use.

Determination of Protein Concentration

Microsomal brain and liver proteins were determined by the method of Lowry et. al., using bovine serum albumin as a standard (26).

Determination of pNP-hydroxylase Activity

Microsomal CYP2E1 (p-Nitrophenol-2-hydroxylase) activity was assayed as described by Reinke and Moyer (27). This enzyme activity was determined with CYP2E1 hydroxylates pNP, its 2-hydroxylation to form of 4-Nitrocatechol was measured spectrophotometrically.

Reaction Medium

1 ml of reaction medium contains microsomal protein, as a substrate pNP, 400 mM pH 6.8 tris buffer, 100 mM glucose-6-phosphate, as a cofactor 20 mM NADP⁺, 100 mM MgCl₂, 400 mM pH 7.8 potassium phosphate buffer and 500 U/0.15 ml glucose-6-phosphate dehydrogenase.

Procedure

The reaction medium (tris buffer, pNP and microsomal protein, respectively) was added to the each test tube. Test tubes containing all substance in the reaction medium were taken in a shaking water bath at 37 °C. Reaction was initiated with the addition of NADPH generating system to each test tube. The samples incubated at 37 °C for 30 min. in a shaking water bath. At the end of the incubation, each reaction was stopped by the addition of 500 µl of 0,75 N (v/v) perchloride acid. Centrifuged for 45 min. at 5 000 g to remove denatured proteins in tubes. After centrifugation, 1 ml of supernatant was taken into another tube. 250 µl of 2 M NaOH was added to complete ionization of 4-Nitrocatechol which formed in the reaction. After the color change

observed in the tubes, centrifuged 5 000 g for 15 min. for the removal of NaOH precipitates. Immediately, after the last centrifugation, the absorbance values were read at a wavelength of 546 nm spectrophotometrically.

RESULTS AND DISCUSSION

In this study, optimum conditions for CYP2E1 activity in C57Bl/6 mouse liver and brain microsomes were investigated with respect to protein and p-nitrophenol concentration and incubation time.

Effect of Protein Amount on CYP2E1 Activity

The effect of protein amount on CYP2E1 activity in brain was examined in 1 ml incubation medium by the addition of 1.0 mg, 2.0 mg, 4.0 mg and 6.0 mg protein. Increasing CYP2E1 activity was observed to be linear in the range of 1.0-4.0 mg protein. Maximum activity of CYP2E1 was measured at 4.0 mg protein in the brain. While examining the effect of protein amount on hepatic CYP2E1 activity in mice, in the incubation medium was studied by the addition of 1.0 mg, 2.0 mg, 3.0 mg and 4.0 mg protein. Maximum activity was observed at 3.0 mg (Table 1, Figure 1).

Table 1. Results of the optimization of protein amount on CYP2E1 activity in C57Bl/6 brain and liver*.

Amount of Protein (mg)	CYP2E1 Enzyme Activity (nmol 4-nitro catechol/min.)	
	Brain	Liver
1.0 mg	0,024 ± 0,004	0,121 ± 0,008
2.0 mg	0,050 ± 0,007	0,241 ± 0,014
3.0 mg	-	0,331 ± 0,011
4.0 mg	0,116 ± 0,008	0,134 ± 0,016
6.0 mg	0,050 ± 0,005	-

* All the values are mean ± SEM of 3 experiments. Each experiment was conducted duplicate.

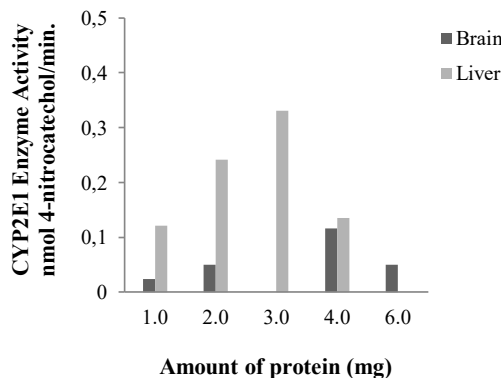


Figure 1. Effect of protein amount on CYP2E1 activity in C57Bl/6 mouse brain and liver.

Effect of Incubation Time on CYP2E1 Activity

The effect of incubation time on CYP2E1 activity in brain and liver was tested with different incubation time between 10-60 min. and maximum activities were observed at 30 minutes in both brain and liver microsomes (Table 2, Figure 2).

Table 2. Results of the optimization of incubation time on CYP2E1 activity in C57Bl/6 brain and liver*.

Incubation Time (min.)	CYP2E1 Enzyme Activity (nmol 4-nitro catechol/mg)	
	Brain	Liver
10 min.	0,222 ± 0,065	1,93 ± 0,206
20 min.	0,484 ± 0,050	2,74 ± 0,180
30 min.	0,800 ± 0,089	4,08 ± 0,191
60 min.	0,406 ± 0,099	2,43 ± 0,262

* All the values are mean ± SEM of 3 experiments. Each experiment was conducted duplicate.

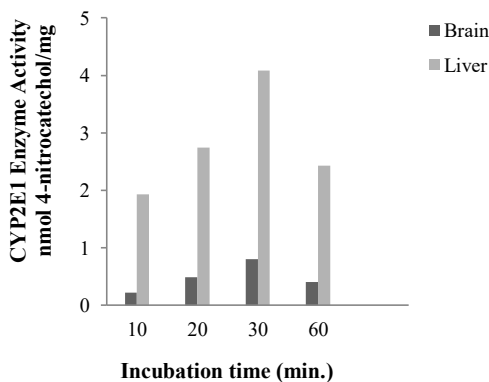


Figure 2. Effect of incubation time on CYP2E1 activity in C57Bl/6 brain and liver.

Effect of Substrate Concentration on CYP2E1 Activity

The effect of pNP concentration on CYP2E1 activity in brain was examined in 1 ml reaction medium by the addition of 62,5 μM, 125 μM, 187,5 μM and 250 μM of pNP. Increasing CYP2E1 activity was observed to be linear in the range of 62,5 μM-125 μM of pNP and maximum activity was observed at 125 μM of pNP in brain. The effect of pNP concentration on CYP2E1 activity in liver was examined in 1 ml reaction medium by the addition of 25 μM, 62,5 μM, 125 μM and 250 μM of pNP and maximum activity was observed at 125 μM of pNP in liver (Table 3, Figure 3). Some studies have reported that CYP2E1 activity was inhibited at concentrations higher than 200 μM of pNP (10,28,29) and these studies lends support to our findings.

Table 3. Results of the optimization of substrate concentration on CYP2E1 activity in C57Bl/6 brain and liver*.

p-Nitrophenol Concentration (μM)	CYP2E1 Enzyme Activity (nmol 4-nitrocatechol/mg/min)	
	Brain	Liver
25 μM	-	0,045 \pm 0,005
62,5 μM	0,008 \pm 0,001	0,080 \pm 0,006
125 μM	0,028 \pm 0,001	0,150 \pm 0,004
187,5 μM	0,014 \pm 0,002	-
250 μM	0,006 \pm 0,002	0,085 \pm 0,008

* All the values are mean \pm SEM of 3 experiments. Each experiment was conducted duplicate.

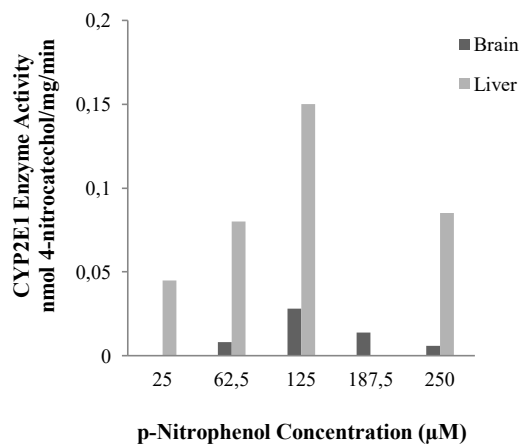


Figure 3. Effect of substrate concentration on CYP2E1 activity in C57Bl/6 brain and liver.

Consequently, in this optimization study it is determined that different reaction conditions are required for CYP2E1 activities in mouse brain and liver microsomes. Furthermore, this study was performed on C57Bl/6 mice which are quite sensitive species. The present results may help to understanding of the functional roles of brain and liver CYP2E1 in prevention and treatment of many diseases.

REFERENCES

- Lewis, D. F. V. (2001). A guide to cytochromes P450 structure and function. Taylor&Francis, London.
- Ferguson, C. S., Tyndale, R. F. (2011). Cytochrome P450 enzymes in the brain: emerging evidence of biological significance. *Trends Pharmacol. Sci.*, 32: 708-714.
- Miksys, S. L., Tyndale, R. F. (2002). Drug-metabolizing cytochrome P450s in the brain. *J. Psychiatry Neurosci.*, 27: 406-415.
- Guengerich, F. P. (1995). Human cytochrome P450 enzymes. In *Cytochrome P450: Structure, Mechanism and Biochemistry*, pp. 473-535, Ortiz de Montellano P. R. (ed.), Plenum, New York.
- Bertz, R. J., Granneman, G. R. (1997). Use of in vitro and in vivo data to estimate the likelihood of metabolic pharmacokinetic interactions. *Clin. Pharmacokinetics*, 32: 210-258.
- Guengerich, F. P., Shimada, T. (1991). Oxidation of toxic and carcinogenic chemicals by human cytochrome P450 enzymes. *Chem. Res. Toxicol.*, 4: 391-407.
- Yamazaki, H., Guo, Z., Guengerich, F. P. (1995). Selectivity of cytochrome P4502E1 in chlorzoxazone 6-hydroxylation. *Drug Metab. Dispos.*, 23: 438-440.
- Lieber, C. S., Rubin, E., DeCarli, L. M. (1970). Hepatic microsomal ethanol oxidizing system (MEOS): differentiation from alcohol dehydrogenase and NADPH oxidase. *Biochem. Biophys. Res. Commun.*, 40: 858-865.
- Porubsky, P., Meneely, M., Scott, E., (2008). Structures of human cytochrome P450 2E1. *J. Biol. Chem.*, 283 : 33698-33707.
- Collom, S. L., Laddusaws, R. M., Burch, A. M., Kuzmic, P., Miller, G. P. (2008). CYP2E1 Substrate Inhibition. Mechanistic interpretation through an effector site for monocyclic compounds. *J. Biol. Chem.*, 283: 3487-3496.
- Cederbaum, A., Wu, D., Mari, M., Bai, J. (2001). CYP2E1-dependent toxicity and oxidative stress in HepG2 cells. *Free Radic. Biol. Med.*, 31: 1539-1543.
- Nanji, A.A., Zhao, S., Sadrzadeh, S. M., Dannenberg, A. J., Tahan, S. R., Waxman, D. J. (1994). Markedly enhanced Cytochrome P450 2E1 induction and lipid peroxidation is associated with severe liver injury in fish oil-ethanol fed rats. *Alcohol Clin. Exp. Res.*, 18: 1280-1285.
- Barnett, C. R., Rudd, S., Flatt, P. R., Ioannides, C. (1993). Sex-differences in the diabetes-induced modulation of rat hepatic cytochrome-P450 proteins. *Biochem. Pharmacol.*, 45: 313-319.
- Raucy, J. L., Lasker, J. M., Kraner, J. C., Salazar, D. E., Lieber, C. S., Corcoran, G. B. (1991). Induction of cytochrome P450IIE1 in the obese overfed rat. *Mol. Pharmacol.*, 39: 275-280.
- Johansson, I., Ekstrom, G., Scholte, B., Puzycki, D., Jomwall, H., Ingelman-Sundberg, M. (1988). Ethanol fasting and acetone-inducible cytochromes P-450 in rat liver: regulation and characteristics of enzyme belonging to the IIB and IIE gene subfamilies. *Biochem.*, 27: 1925-1934.
- Lieber, C. S. (1999). Microsomal ethanol-oxidizing system (MEOS): the first 30 years (1968-1998)- a review. *Alcohol Clin. Exp. Res.*, 23: 991-1007.
- Ravindranath, V., Anandatheerthavarada, H.K. (1989). High activity of cytochrome P450 linked aminopyrene N-demethylase in mouse brain microsomes and associated sex related differences. *Biochem. J.*, 261: 769-773.
- Roberts, B. J., Shof, S. E., Jeong, K. S., Song, B. J. (1994). Induction of CYP2E1 in liver, kidney, brain and intestine during chronic ethanol administration and withdrawal: evidence that CYP2E1 possesses a rapid phase half-life of 6 h or less. *Biochem. Biophys. Res. Commun.*, 205: 1064-1071.
- Warner, M., Gustafsson, J. A. (1994). Effect of ethanol cytochrome P450 in the rat brain. *Proc. Natl. Acad. Sci.*, 91: 1019-1023.
- Parmar, D., Dayal, M., Seth, P. K. (2003). Expression of cytochrome P450s in brain: Physiological, pharmacological and toxicological consequences. *Proceedings of Indian National Academy of Sciences*, 6: 905-928.
- Anandatheerthavarada, H. K., Shankar, S. K., Bhamre, S., Boyd, M. R., Song, B. J., Ravindranath, V. (1993). Induction of cytochrome P-450 IIE1 by chronic ethanol treatment. *Brain Res.*, 601: 279-285.
- Upadhyaya, S. C., Tirumalai, P. S., Boyd, M. R., Mori, T., Ravindranath, V. (2000). Cytochrome P4502E1 in brain: constitutive expression, induction by ethanol and localization by fluorescence in situ hybridization. *Arch. Biochem. Biophys.*, 373: 23-24.
- Park, J. Y., Harris, D. (2003). Construction and assessment of models of CYP2E1: predictions of metabolism from docking, molecular dynamics and density functional theoretical calculations. *J. Med. Chem.*, 46: 1645-1660.

24. Blevins, M., (2008). Human Cytochrome P450 2E1: Functional Comparison to Cytochromes P450 2A13 and 2A6. Graceland University, Master Thesis.
25. Lewis, V. J., Przedborski, S. (2007). Protocol for the MPTP mouse model of Parkinson's disease. *Nature Protocols*, 2: 141-151.
26. Lowry, O. H., Rosebrough, N. J., Farr, A. L., Randall, R. J. (1951). Protein measurement with the folin phenol reagent. *J. Biol. Chem.*, 193: 265-275.
27. Reinke, L. A., Moyer, M. J. (1985). p-Nitrophenol hydroxylation: a microsomal oxidation which is highly inducible by ethanol. *Drug Metab. Dispos.*, 13: 548-552.
28. Koop, D. R. (1986). Hydroxylation of p-nitrophenol by rabbit ethanol-inducible cytochrome P-450 isoenzyme 3a. *Mol. Pharmacol.*, 29: 399-404.
29. Larson, J. R., Coon, M. J., Porter, T. D. (1991). Purification and properties of a shortened form of cytochrome P-450 2E1: deletion of the NH₂-terminal membrane-insertion signal peptide does not alter the catalytic activities. *Proc. Natl. Acad. Sci.*, 88: 9141-9145.